

## ANATOMICAL STUDIES OF SOME HYDROPHYTES COLLECTED FROM DISTRICT BHIMBER, AZAD JAMMU KASHMIR, PAKISTAN

MUHAMMAD RIZWAN<sup>1</sup>, UZMA HANIF<sup>1</sup>, WASIM ABBAS<sup>1</sup>, ALI HASNAIN<sup>1</sup>, SYED KHIZAR SULTAN<sup>1</sup>, GHULAM FATIMAH<sup>1</sup>, AMANAH MEHREEN<sup>1</sup>, TANTRI DYAH AYU ANGRRAENI<sup>3</sup>, MUHAMMAD UMER FAROOQ AWAN<sup>1\*</sup>, MATIBA TUFAIL, NAVEED ANJUM<sup>1</sup>, MUHAMMAD USAMA AZIZ<sup>1</sup>, MUHAMMAD SHOIB<sup>1</sup>

<sup>1</sup>Department of Botany, Government College University, Lahore, Pakistan.

<sup>2</sup>Indonesian Sweetener and Fiber Crops Research Institute, Raya Karangploso, Malang, East Java, Indonesia

Corresponding Author: \*dr.umerfarooqawan@gcu.edu.pk

Received on: 17-09-24; Reviewed on: 11-05-25; Accepted on: 20-05-2025; Published on: 15-06-2025

### Abstract

Hydrophytes play a crucial role in maintaining the ecological balance in aquatic ecosystems. They are bioindicators for water quality, pollution and eutrophication, ultimately assisting habitat restoration. The unique anatomical features of hydrophytes are essential for their survival in waterlogged or submerged environments. Studying these traits reveal how plants evolve to overcome challenges like, low oxygen availability, nutrient absorption and reduced mechanical support. The present study was carried out for the anatomical study of hydrophytes collected from district Bhimber, Azad Jammu and Kashmir, Pakistan. Total three species, *Hydrilla verticillata* (L.f.) Royle, *Eichhornia crassipes* (Mart.) Solms and *Nymphaea alba* L. were collected and preserved in the fixative solutions and subjected to anatomical characterization using a light microscope. In the T.S. of the stem of *Nymphaea alba* L., trichomes were observed ( $L=4.50\pm0.7$ ,  $W=2.11\pm0.48$ ) profusely surrounding the layer of epidermis. Larger aerenchyma was observed ( $L=121.50\pm47.34$ ,  $W=92.76\pm32.26$ ). In the T.S of *Hydrilla verticillata* (L.f.) Royle, larger aerenchyma cells were also observed ( $L=41.84\pm8.86$ ,  $W=29.49\pm2.1$ ). In the T.S of *Eichhornia crassipes* (Mart.) Solms aerenchyma was larger among other cells ( $L=40.50\pm1.00$ ,  $W=28.25\pm1.48$ ). Trichomes were also examined ( $L=3.46\pm1.06$ ,  $W=1.73\pm0.20$ ). Larger aerenchyma and multicellular trichomes were observed, indicating that these species are better adapted to the influx of excessive water in their aquatic habitats. Overall, the research is correlating the anatomical characterization of the hydrophytes with their environmental and climatic factors helping to understand the mechanism of ecological adaptations to their aquatic environment.

**Keywords:** Hydrophytes, anatomical study, climatic factors, Trichomes.

### INTRODUCTION:

Plant species are commonly classified at the ecological level according to their evolutionary adaptations, phenotypic variations, and the interplay between the evolution of their morphological, physiological, and anatomical traits in response to their environmental conditions. Hydrophytes establish specialized ecological roles in many aquatic environments, including freshwater lakes, marshy areas along the peripheries of the coastal regions, wetlands, ponds and inland marshes (Rodrigo, 2021). Hydrophytes demonstrate a diverse range of phenomenal adaptations that facilitate them to flourish and thrive within aquatic environments (Tiner, 2006). These adaptations encompass a range of anatomical, physiological, and morphological changes that are directed at enhancing their

efficiency of nutrition absorption, regulation of buoyancy, photosynthesis, and exchange of gases in order to effectively respond to the demands posed by aquatic environments (Ronzhina *et al.*, 2001). Hydrophytes are magnificently essential and are widely used in restorative practices wetland ecosystems (Zhao *et al.*, 2016). The examination of the anatomical characteristics of various morphological components of hydrophytes facilitates a comprehensive understanding of their adaptive strategies and enables a more thorough and insightful analysis of the mechanisms they employ to thrive in aquatic environments (Jayeola and Folorunso, 2009). Anatomical analysis is a key factor for the identification of different plant species. In order to study plant systematics, anatomical

analysis is pragmatically necessary. Most of the time, with the change of plant habitat, a change in the epidermal size, type of the stomata, trichomes, and subsidiary cells can be observed (Werker, 2000).

The present research aimed to investigate anatomical features of morphological components of some hydrophytes collected from the district Bhimber, Azad Jammu and Kashmir, Pakistan, as previously no published data is available for the anatomical features of hydrophytes of Azad Jammu and Kashmir. The biodiversity found in Azad Jammu and Kashmir is unrivalled as compared to other regions of Pakistan (Ajaib *et al.*, 2012). Bhimber is a well-recognized district that shares a border with Punjab, Pakistan to the south Bhimber is situated in the southern region of Azad Jammu and Kashmir, while its eastern border is adjacent to the Indian-administered Kashmir. The study area (District: Bhimber, Azad Jammu and Kashmir) is located between latitude 32-48° to 33-34° and longitude 73-55° to 74-45°. The total area of district Bhimber is 1516 square kilometers with an altitude of 1118 feet above sea level (Maqbool *et al.*, 2019). District Bhimber exhibits a diverse geography, hydrophytes (I.e. *Nymphaea alba* L., *Hydrilla verticillata* (L.f.) Royle, and *Eichhornia crassipes* (Mart.) Solms) in different aquatic habitats (ponds, ditches, freshwater lakes, and marshes).

*Nymphaea* is a genus of aquatic plants commonly referred as water lilies. There are many species of *Nymphaea* that are distributed worldwide, with variations in their morpho-anatomical characteristics, range of distribution, and morphological characteristics of flower (i.e. size, colour, shape, arrangement, and differences in the number of sepals, petals, and stamens), Morpho-anatomical characteristics of their leaves (i.e. variation in size, shape, and color of the leaves; the shape of the leaves blade, venation pattern, and texture of the surface of their leaves), and

morphological features of their roots or rhizome (Harrison, 1955).

The flowers are the most striking feature of *Nymphaea alba* L. They are large, fragrant, and typically have pure white petals that radiate outward from a yellow centre. Each flower can measure up to 12-15 cm in diameter, making it highly visible in the aquatic environment (Wheeler, 1997) *Nymphaea alba* L. plays a crucial role in improving water quality. Its extensive root system absorbs excess nutrients like nitrogen and phosphorus, which helps prevent water eutrophication and the growth of harmful algae (Nierbauer *et al.*, 2014). The stem of *Nymphaea alba* L. has abundant aerenchyma observed along with the presence of multicellular trichomes (Bavaru and Rodica, 2002).

*Hydrilla verticillata* (L.f.) Royle, commonly known as Water thyme. It is a highly adaptable and invasive aquatic plant that belongs to the Hydrocharitaceae family (Pieterse, 1981). It is native to some warmer areas of Asia, Europe, Africa, the USA, and Australia but has become a notorious invasive species in many regions across the world (Langeland, 1996).

*Hydrilla verticillata* (L.f.) Royle is characterized by long, slender stems that can grow up to 25 feet in length (Yeo *et al.*, 1984). The leaves are arranged in whorls, along the stem. They have serrated and narrow leaves, resembling those of thyme (Verkleij *et al.*, 1983; Sumithran *et al.*, 2013). *Hydrilla verticillata* (L.f.) Royle is highly adaptable to a wide range of aquatic habitats. It can thrive in both stagnant and flowing water, and it tolerates various water temperatures and depths (Michel *et al.*, 2004). It is immensely tolerant to a wide range of pH. It is adapted to carry out the process of photosynthesis even in lower-intensity sunlight (Steward, 1993). They can reproduce vegetatively so they tremendously are fast acting in competing with

other aquatic plants and aggressively colonize wetland ecosystems (Langeland, 1996). It can reduce water clarity, block sunlight penetration, and deplete oxygen levels, which can harm native aquatic species, including fish and invertebrates (Jones *et al.*, 2003).

*Eichhornia crassipes* (Mart.) Solms, commonly known as Water hyacinth, is a free-floating aquatic plant native to south America but has become a notorious aquatic weed due to its invasive nature and widely spread across different regions of the world (Milne *et al.*, 2006). Water hyacinth is recognized by its attractive appearance, featuring bright green, glossy, oval-shaped leaves and beautiful lavender or violet flowers with a yellow spot. This plant thrives in slow-moving or stagnant freshwater bodies such as ponds, lakes, rivers, and canals. Water hyacinths may widely spread due to vegetative means of reproduction and have a rapid growth rate (Coetzee *et al.*, 2009). The rapid growth of water hyacinth can lead to the formation of thick mats that cover the surface of the water. This blocks sunlight, which can harm submerged aquatic plants and disrupt the food web (Ayanda *et al.*, 2020).

*Eichhornia crassipes* (Mart.) Solms are widely employed for phytoremediation (Xia and Ma, 2006) and have enormous tendencies to absorb and accumulate heavy metals and toxic pollutants from the water bodies (Jafari, 2010).

This anatomical analysis of hydrophytes is pragmatically essential to understanding the

complex morphological, anatomical, and physiological adaptations to survive in aquatic habitats.

Moreover, the present study aims to systematically document and classify hydrophyte species based on their anatomical attributes present in district Bhimber, Azad Kashmir, while also providing valuable perspectives on their evolutionary connections and ecological functions within the indigenous environment.

## MATERIALS AND METHODS

### Collection of Samples:

The specimens were hydrophytes collected from the marshy areas, ditches, ponds, moist regions, and wet grasslands of the district Bhimber AJK and got authenticated by Dr. Uzma Hanif, Department of Botany, GC University Lahore, Pakistan (Table 1). The specimens were collected and preserved in the fixative solutions as described by Jain and Rao (1977). The species were identified morphologically by using standard methods of flora, dried up and mounted on herbarium sheets. These plants were deposited and got voucher number from Department of Botany, Government College University, Lahore.

### Preparation of Reagents:

#### Fixative Solution:

Leaf tissue was fixed according to the methodology described by Bomblies *et al.* (2008). The Fixative solution was prepared by mixing 500ml 95% ethanol, 100 mL 14% formalin, 50 mL Acetic acid, and the final volume rose up to 1L.

**Table 1. Collection of Species from Bhimber AJK**

Serial Number	Name of Species	Collection Site	Date of Collection
01	<i>Nymphaea alba</i> L.	Bhimber AJK	November 27, 2022
02	<i>Hydrilla verticillata</i> (L.f.) Royle	Bhimber AJK	November 27, 2022
03	<i>Eichhornia crassipes</i> (Mart.) Solms	Bhimber AJK	November 27, 2022

**Preparation of Reagents:****Fixative Solution:**

Leaf tissue was fixed according to the methodology described by Bomblies *et al.* (2008). The Fixative solution was prepared by mixing 500 ml 95% ethanol, 100 mL 14% formalin, 50 mL Acetic acid, and the final volume rose up to 1L.

**Preparation of Toluidine Blue Stain:**

Toluidine blue stain was prepared according to method described by Bergholt *et al.* (2019). 0.5g toluidine blue was dissolved in a small amount of distilled water and then rose the final volume up to 100ml by adding distilled water.

**Anatomical Study of Stem:**

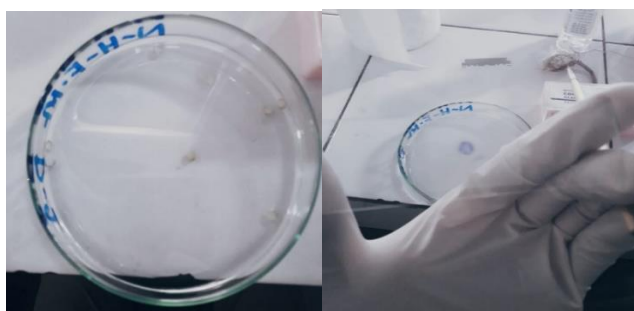
The following steps were performed to study the anatomical characteristics:

**Section cutting procedure:**

The section-cutting procedure was performed as described by (Zelko *et al.*, 2012).

**Toluidine Blue Staining Procedure:**

For toluidine blue staining, the following steps were performed according to the method described by Lux *et al.* (2005). The sections were placed in 70% alcohol for approximately 1 minute prior to stain. Then these sections were placed on a glass slide by using a camel hairbrush. The fine sections placed on the glass slide were covered with the coverslips and then observed under the light microscope.

**Plate 1: Section cutting and toluidine blue staining****Micrometry and Microphotography:**

Digital photographs were taken by and the measurements of different cells were carried out by

using standard methods of micrometry as described by Quesnel (1971).

**RESULTS**

### Anatomical Analysis of the Transverse Sections of Stem of *Nymphaea alba* L.

Stem sections of *Nymphaea alba* L. were examined under the light microscope; a layer of epidermis was clearly observed surrounded by trichomes. The glandular trichomes were profusely surrounding the layer of the epidermis (Plate 2B). Length of the trichomes was observed as  $4.50 \pm 0.77$  ( $\mu\text{m}$ ) ranging from  $3.85 \mu\text{m}$  to  $5.36 \mu\text{m}$ . However, the width of the trichomes was observed as  $2.11 \pm 0.48$  ( $\mu\text{m}$ ) ranging from  $1.56 \mu\text{m}$  to  $2.48 \mu\text{m}$  in different trichomes (Table 2). While studying the *Nymphaea alba* L., aerenchyma was clearly observed. The presence of aerenchyma signified the anatomical adaptation to absorb the excess amount of water in their aquatic habitat (Plate 2B). The length of the aerenchyma was observed as  $121.50 \pm 47.34$  ( $\mu\text{m}$ ) ranging from  $71.03 \mu\text{m}$  to  $164.94 \mu\text{m}$  in different aerenchyma. However, the width of the aerenchyma was observed as  $92.76 \pm 32.26$  ( $\mu\text{m}$ ) ranging from  $70.43 \mu\text{m}$  to  $129.76 \mu\text{m}$  in different aerenchyma in the stem of *Nymphaea alba* L. (Table 2). Small vascular bundles were observed in a layer beneath the epidermis (Plate 2B). Length of the small vascular bundles was observed as  $7.45 \pm 0.31$  ( $\mu\text{m}$ ) ranging from  $7.26 \mu\text{m}$  to  $7.81 \mu\text{m}$ . However, the width of the small vascular bundles was observed as  $92.76 \pm 32.26$  ( $\mu\text{m}$ ) ranging from  $70.43 \mu\text{m}$  to  $129.76 \mu\text{m}$  (Table 4.1). The larger vascular bundle was also observed. The length of the larger vascular bundle was observed as  $54.24 \pm 5.06$  ( $\mu\text{m}$ ) ranging from  $51.16 \mu\text{m}$  to  $60.08 \mu\text{m}$ . However, the width of the larger vascular bundle was measured as  $59.48 \pm 1.29$  ( $\mu\text{m}$ ) ranging from  $58.00 \mu\text{m}$  to  $60.42 \mu\text{m}$  (Plate 2C, Table 2).

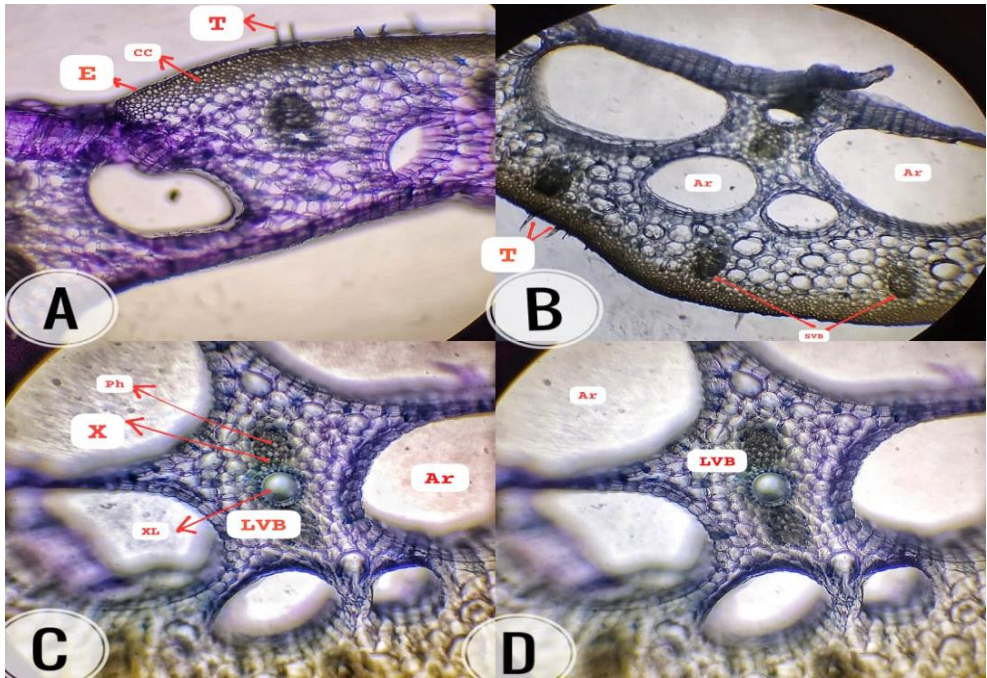
Xylem and Phloem were clearly observed in the larger vascular bundles (Plate 2C). Length of the xylem was measured as  $17.91 \pm 1.62$  ( $\mu\text{m}$ ) ranging from  $16.32 \mu\text{m}$  to  $19.56 \mu\text{m}$ . However, the width of the xylem was measured as  $6.55 \pm 0.79$  ( $\mu\text{m}$ ) ranging

from  $6.04 \mu\text{m}$  to  $7.41 \mu\text{m}$  in different samples of the stem of *Nymphaea alba* L. (Table 2). The phloem was comparatively larger in length and broader in the width than xylem (Figure 2). The length of the phloem was measured as  $29.57 \pm 1.97$  ( $\mu\text{m}$ ) ranging from  $27.62 \mu\text{m}$  to  $31.56 \mu\text{m}$ . However, the width of the phloem was observed as  $31.47 \pm 2.54$  ( $\mu\text{m}$ ) ranging from  $29.07 \mu\text{m}$  to  $34.14 \mu\text{m}$  in the phloem of different sections of the stem of *Nymphaea alba* L. (Table 2). The length of the collenchyma cells was measured as  $40.36 \pm 6.67$  ( $\mu\text{m}$ ) ranging from  $33.62 \mu\text{m}$  to  $46.96 \mu\text{m}$ . The width of the collenchyma cells was measured as  $28.85 \pm 9.89$  ( $\mu\text{m}$ ) ranging from  $17.80 \mu\text{m}$  to  $36.88 \mu\text{m}$  (Plate 2A, Table 2).

### Anatomical Analysis of the Transverse Sections of the Stem of *Hydrilla verticillata* (L.f.) Royle

In the transverse section of the stem of *Hydrilla verticillata* (L.f.) Royle, a compact arrangement of the cells was observed and larger aerenchyma cells were also observed (Plate 3). However, different cells and tissues such as parenchyma, ground tissues, xylem, and phloem were present and observed using the light microscope at 40X. Collenchyma cells were observed (Plate 3D). Length of the collenchyma cells was measured as  $36.93 \pm 2.39$  ( $\mu\text{m}$ ) ranging from  $34.31 \mu\text{m}$  to  $39.00 \mu\text{m}$ . However, the width of the collenchyma cells was measured as  $12.94 \pm 0.54$  ( $\mu\text{m}$ ) ranging from  $12.39 \mu\text{m}$  to  $13.48 \mu\text{m}$  (Table 3). Length of the aerenchyma cells was measured as  $41.84 \pm 8.86$  ( $\mu\text{m}$ ) ranging from  $34.29 \mu\text{m}$  to  $51.61 \mu\text{m}$ . However, the width of the aerenchyma cells was observed as  $29.49 \pm 2.1$  ( $\mu\text{m}$ ) ranging from  $27.35 \mu\text{m}$  to  $31.67 \mu\text{m}$  (Plate 3 A, Table 3). Vascular bundles were present in which the xylem and phloem were clearly observed (Plate 3C). Length of the xylem was observed as  $21.78 \pm 2.16$  ( $\mu\text{m}$ ) ranging from  $19.34 \mu\text{m}$  to  $23.45 \mu\text{m}$ . The width of the xylem was measured as  $14.37 \pm 0.94$  ( $\mu\text{m}$ ) ranging from  $13.45 \mu\text{m}$  to  $15.39 \mu\text{m}$  (Table 3). Length of the

phloem was measured as  $34.78 \pm 2.08$  ( $\mu\text{m}$ ) ranging from  $33.14 \mu\text{m}$  to  $37.13 \mu\text{m}$ . However, the width of the phloem was observed as  $13.67 \pm 1.00$  ( $\mu\text{m}$ ) ranging from  $12.60 \mu\text{m}$  to  $14.58 \mu\text{m}$  (Plate 3C, Table 3).

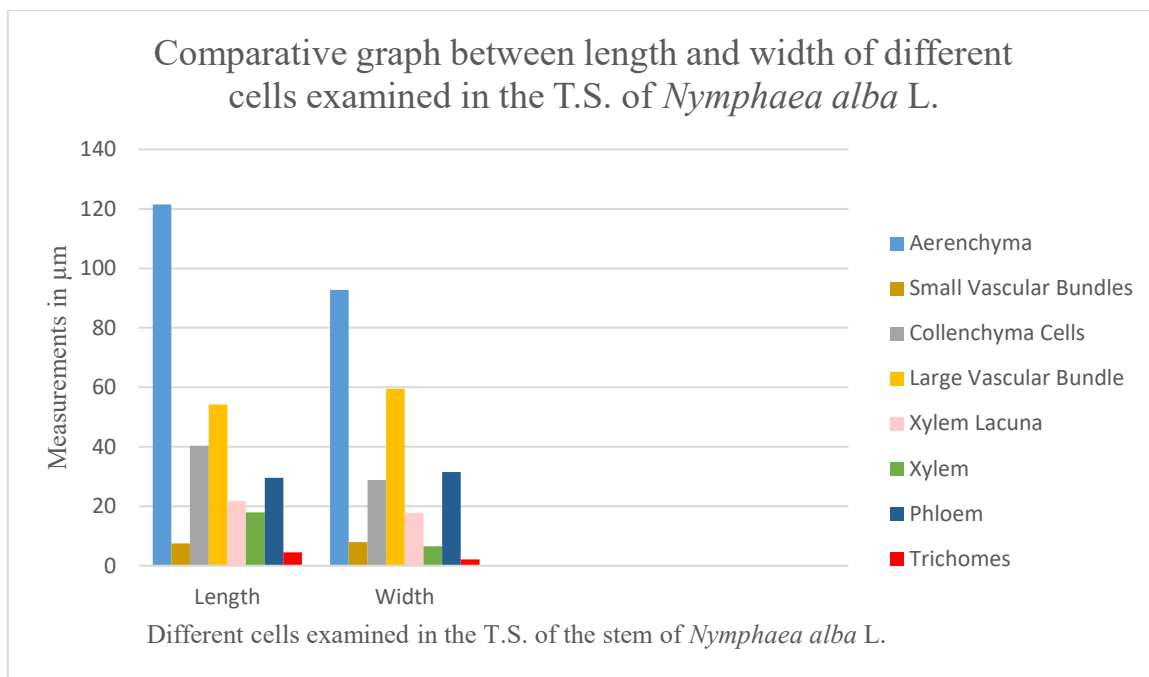


**Plate 2:** T.S. of the stem of *Nymphaea alba* L. A. (E epidermis CC collenchyma cells T trichomes) B. Ar aerenchyma SVB smaller vascular bundles T trichomes) C. LVB larger vascular bundles XL xylem lacuna X xylem Ph phloem) D. (Ar aerenchyma) (40X)

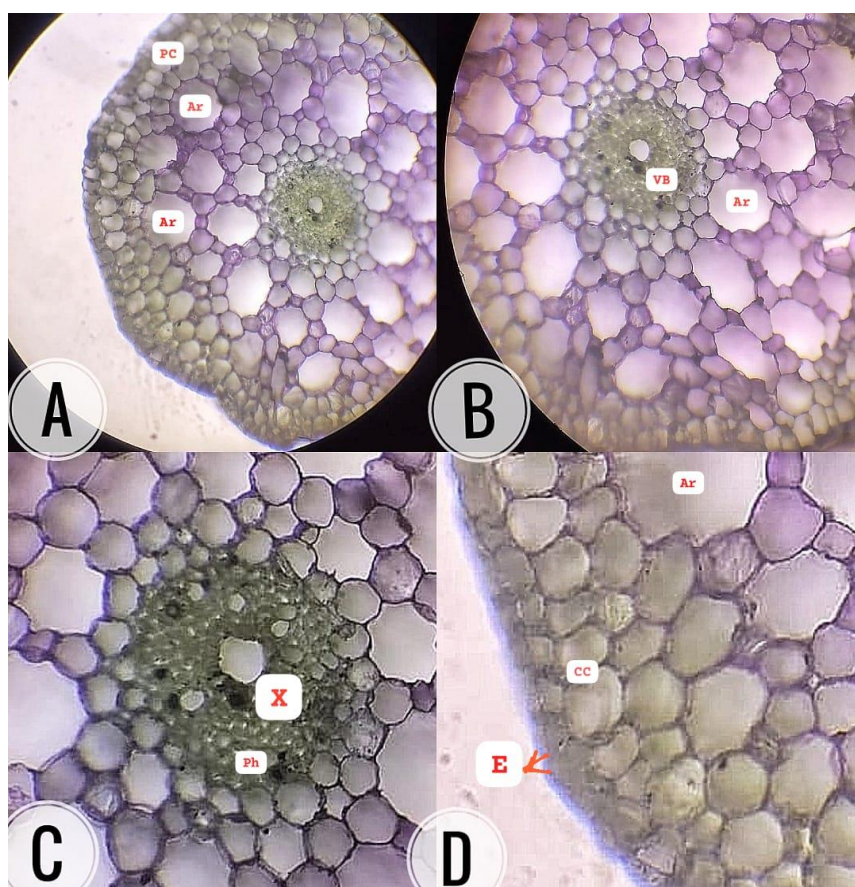
**Table 2:** Length and width of Different Cells in the T.S. of the stem of *Nymphaea alba* L.

Sr. No.	Parameters	Length ( $\mu\text{m}$ )	Range of the length of cells ( $\mu\text{m}$ )	Width ( $\mu\text{m}$ )	Range of the width of cells ( $\mu\text{m}$ )
01	Aerenchyma	$121.50 \pm 47.34$	71.03-164.94	$92.76 \pm 32.26$	70.43-129.76
02	Small vascular bundles	$7.45 \pm 0.31$	7.26-7.81	$7.97 \pm 1.25$	6.71-9.22
03	Collenchyma cells	$40.36 \pm 6.67$	33.62-46.96	$28.85 \pm 9.89$	17.80-36.88
04	Large Vascular Bundle	$54.24 \pm 5.06$	51.16-60.08	$59.48 \pm 1.29$	58.00-60.42
05	Xylem lacuna	$21.84 \pm 6.59$	15.00-28.16	$17.77 \pm 3.60$	14.99-21.84
06	Xylem	$17.91 \pm 1.62$	16.32-19.56	$6.55 \pm 0.79$	6.04-7.41
07	Phloem	$29.57 \pm 1.97$	27.62-31.56	$31.47 \pm 2.54$	29.07-34.14
08	Trichomes	$4.50 \pm 0.77$	3.85-5.36	$2.11 \pm 0.48$	1.56-2.48





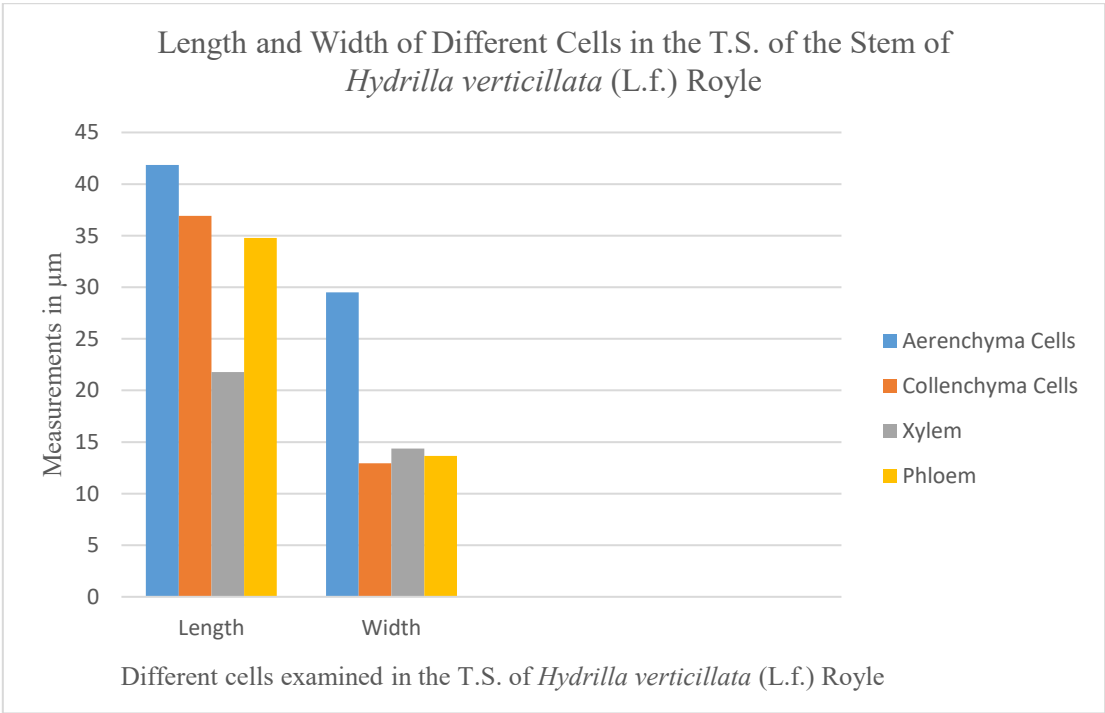
**Figure 1:** Comparative graph between length and width of different cells examined in the T.S. of the stem of *Nymphaea alba* L.



**Plate 3:** T.S. of the stem of *Hydrilla verticillata* (L.f.) Royle **A.** (PC parenchyma cells Ar aerenchyma cells) **B.** (VB vascular bundle) **C.** (X xylem Ph phloem) **D.** (CC collenchyma cells E epidermis cells) (40X)

**Table 3:** Length and width of different cells examined in the T.S. of *Hydrilla verticillata* (L.f.) Royle

Sr. No.	Parameters	Length (μm)	Range of the length of cells (μm)	Width (μm)	Range of the width of cells (μm)
01	Aerenchyma Cells	41.84±8.86	34.29-51.61	29.49±2.1	27.35-31.67
02	Collenchyma Cells	36.93±2.39	34.31-39.00	12.94±0.54	12.39-13.48
03	Xylem	21.78±2.16	19.34-23.45	14.37±0.94	13.45-15.34
04	Phloem	34.78±2.08	33.14-37.13	13.67±1.00	12.60-14.58



**Figure 2:** Comparative graph between length and width of different cells examined in the T.S. of the stem of *Hydrilla verticillata* (L.f.) Royle

**Anatomical Analysis of the Transverse Sections of the Stem of *Eichhornia crassipes* (Mart.) Solms**

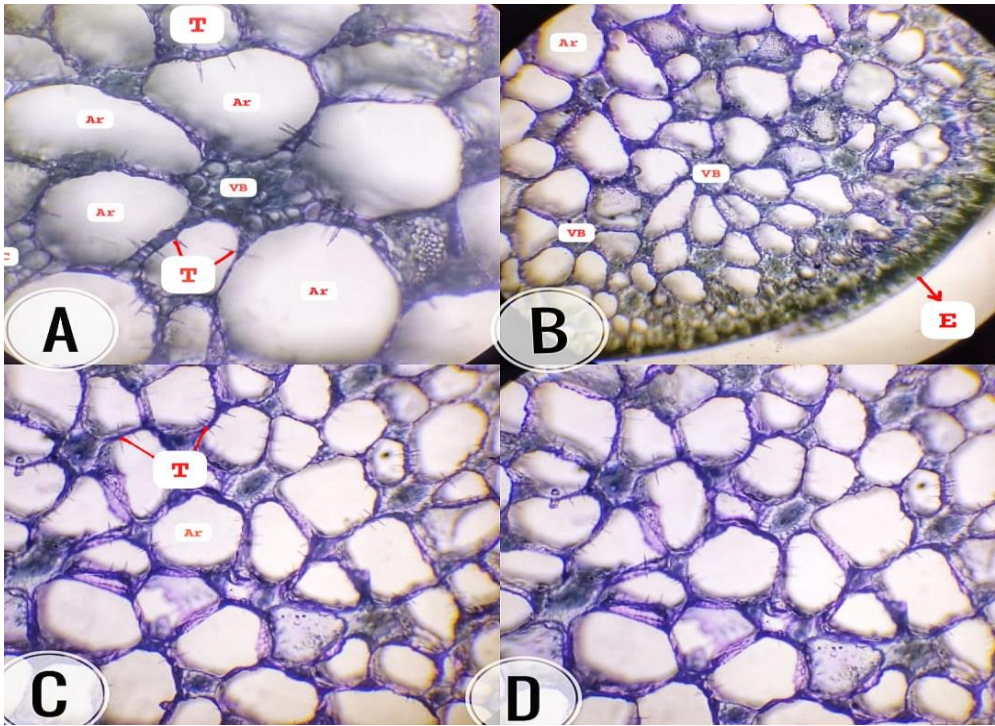
The transverse section of the stem of *Eichhornia crassipes* (Mart.) Solms was observed under the light microscope at 40X. Aerenchyma was abundantly present. Vascular bundles were present

in which xylem and phloem were observed (Plate 4). The length of the aerenchyma cells was measured as 40.50±1.00 (μm) ranging from 39.58μm to 41.57μm. However, the width of the aerenchyma cells was observed as 28.25±1.48 (μm) ranging from 26.56μm to 29.35μm (Plate 4A, Table 4). Vascular



bundles were present in which the xylem and phloem were clearly observed (Plate 4B). Length of the xylem was observed as  $22.98 \pm 6.48$  ( $\mu\text{m}$ ) ranging from  $16.36\mu\text{m}$  to  $29.32\mu\text{m}$ . The width of the xylem was measured as  $17.36 \pm 3.57$  ( $\mu\text{m}$ ) ranging from

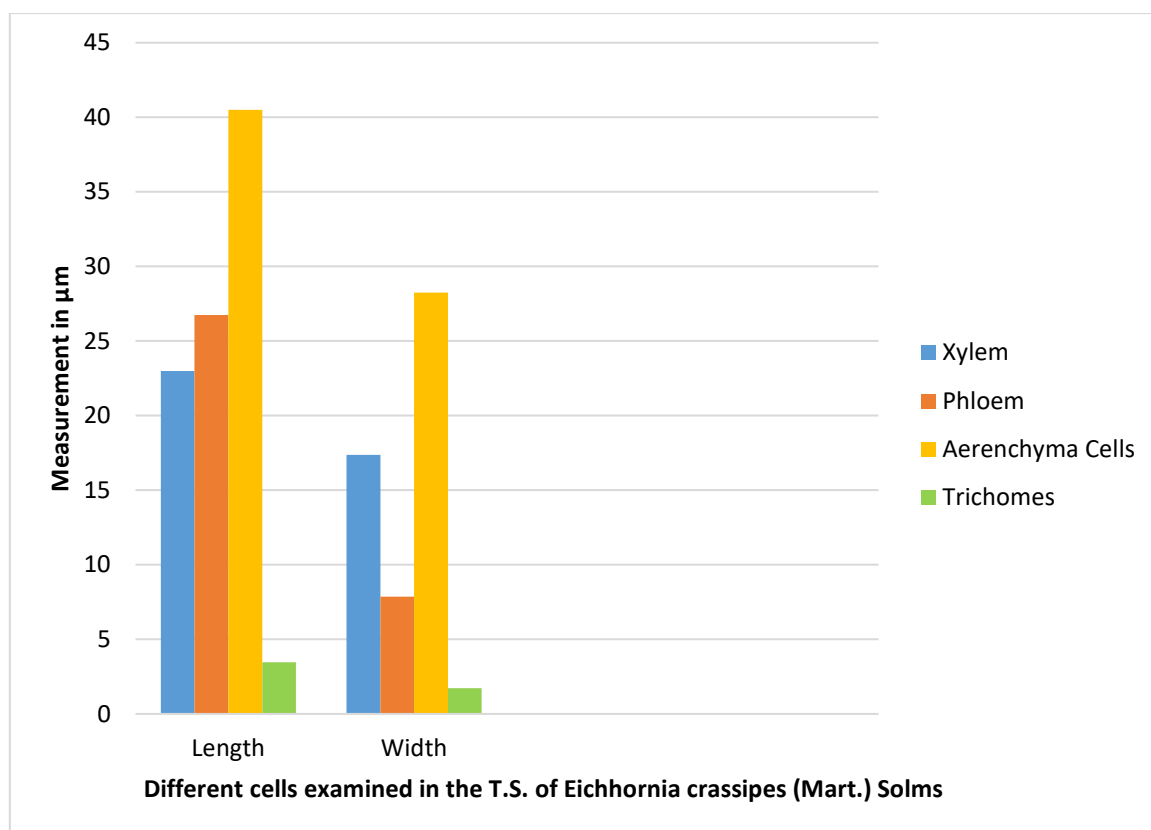
$13.83\mu\text{m}$  to  $20.98\mu\text{m}$ . The length of the phloem was measured as  $26.74 \pm 2.64$  ( $\mu\text{m}$ ) ranging from  $23.92\mu\text{m}$  to  $29.17\mu\text{m}$ . However, the width of the phloem was observed as  $7.86 \pm 0.71$  ( $\mu\text{m}$ ) ranging from  $7.37\mu\text{m}$  to  $8.69\mu\text{m}$  (Table 4).



**Plate 4:** T.S. of the stem of *Eichhornia crassipes* (Mart.) Solms **A.** (Ar aerenchyma cells VB vascular bundle T trichomes) **B.** (E epidermis) **C.** (T trichomes) (40X)

**Table 4:** Length and width of different cells examined in the T.S. of *Eichhornia crassipes* (Mart.) Solms

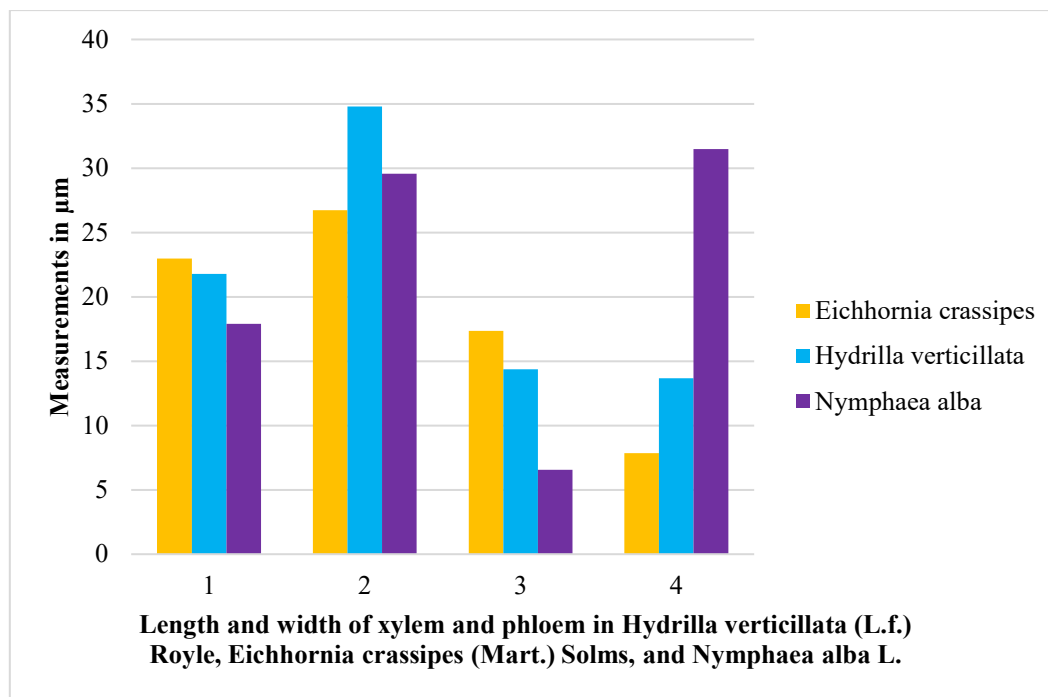
Sr. No.	Parameters	Length ( $\mu\text{m}$ )	Range of the length of cells ( $\mu\text{m}$ )	Width ( $\mu\text{m}$ )	Range of the width of cells ( $\mu\text{m}$ )
01	Aerenchyma Cells	$40.50 \pm 1.00$	39.58-41.57	$28.25 \pm 1.48$	26.56-29.35
02	Vascular Bundle	$38.27 \pm 10.17$	32.20-50.02	$26.79 \pm 1.61$	24.93-27.80
03	Xylem	$22.98 \pm 6.48$	16.36-29.32	$17.36 \pm 3.57$	13.83-20.98
04	Phloem	$26.74 \pm 2.64$	23.92-29.17	$7.86 \pm 0.71$	7.37-8.69
05	Trichomes	$3.46 \pm 1.06$	2.35-4.48	$1.73 \pm 0.20$	1.54-1.94



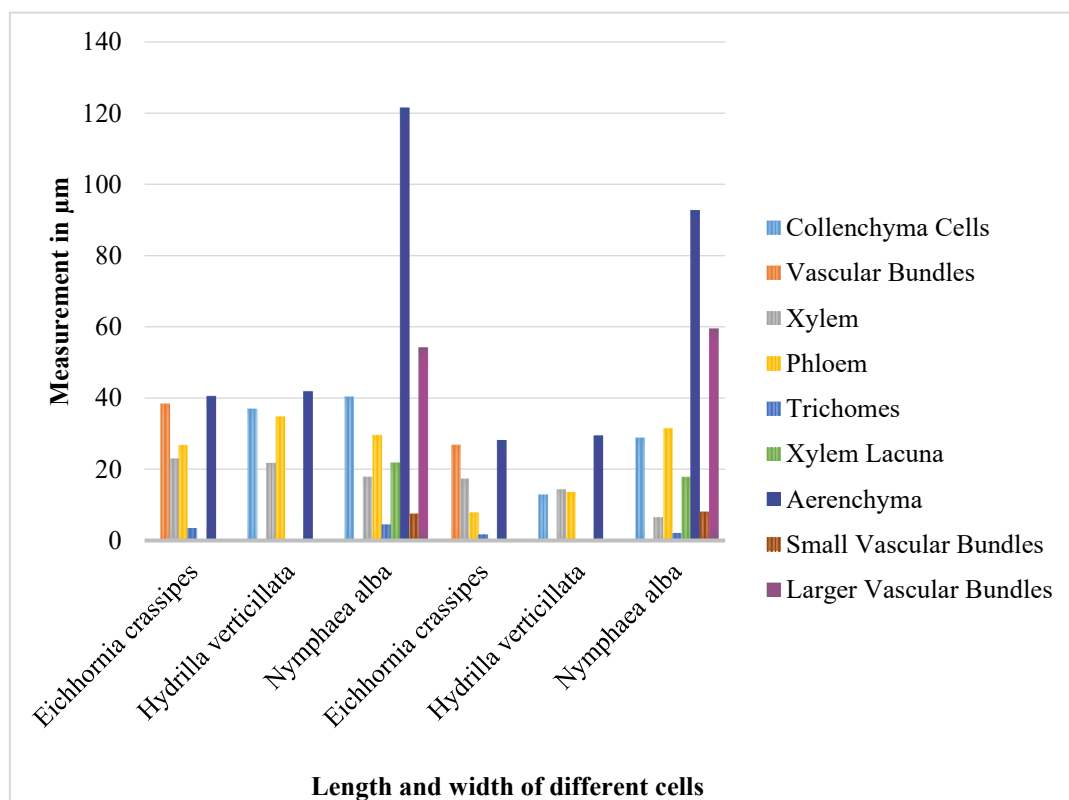
**Figure 3:** Comparative graph between length and width of different cells examined in the T.S. of the stem of *Eichhornia crassipes* (Mart.) Solms

**Table 5:** Comparative anatomical analysis of xylem and phloem in *Hydrilla verticillata* (L.f.) Royle, *Eichhornia crassipes* (Mart.) Solms, and *Nymphaea alba* L.

Sr. No.	Name of the Species	Parameters	Length ( $\mu\text{m}$ )	Width ( $\mu\text{m}$ )
01	<i>Hydrilla verticillata</i> (L.f.) Royle	Xylem	21.78 $\pm$ 2.16	14.37 $\pm$ 0.94
02	<i>Eichhornia crassipes</i> (Mart.) Solms	Xylem	22.98 $\pm$ 6.48	17.36 $\pm$ 3.57
03	<i>Nymphaea alba</i> L.	Xylem	17.91 $\pm$ 1.62	6.55 $\pm$ 0.79
04	<i>Hydrilla verticillata</i> (L.f.) Royle	Phloem	34.78 $\pm$ 2.08	13.67 $\pm$ 1.00
05	<i>Eichhornia crassipes</i> (Mart.) Solms	Phloem	26.74 $\pm$ 2.64	7.86 $\pm$ 0.1
06	<i>Nymphaea alba</i> L.	Phloem	29.57 $\pm$ 1.97	31.47 $\pm$ 2.54



**Figure 4:** Comparative Anatomical Analysis of Xylem and Phloem in *Hydrilla verticillata* (L.f.) Royle, *Eichhornia crassipes* (Mart.) Solms, and *Nymphaea alba* L.



**Figure 5:** Comparative anatomical analysis (length and width) of different cells examined in the T.S. of *Eichhornia crassipes* (Mart.) Solms, *Hydrilla verticillata* (L.f.) Royle, and *Nymphaea alba* L.

## DISCUSSION

Hydrophytes exhibit a wide range of adaptations that facilitate them to grow and thrive within aquatic environments (Tiner, 2006; Rodrigo. 2021). These adaptations encompass a range of anatomical, physiological, and morphological changes that are directed at enhancing their efficiency of nutrition absorption, regulation of buoyancy, photosynthesis, and exchange of gases to effectively respond to the demands posed by aquatic environments (Ronzhina *et al.*, 2001). The examination of the anatomical characteristics of various morphological components of hydrophytes facilitates a comprehensive understanding of their adaptive strategies and enables a more thorough and insightful analysis of the mechanisms they employ to thrive in aquatic environments (Jayeola and Folorunso, 2009).

This research work was conducted to collect different hydrophytes of the district Bhimber, Azad Jammu and Kashmir, Pakistan. The focus of the research was to identify and authenticating the taxonomic status of the collected species, based on the anatomical analysis of their morphological parts. In this research, anatomical features of the transverse sections of the stem of *Eichhornia crassipes* (Mart.) Solms, *Nymphaea alba* L., and *Hydrilla verticillata* (L.f.) Royle have been studied by using light microscope (40X). Three hydrophytes, *Eichhornia crassipes* (Mart.) Solms, *Nymphaea alba* L., and *Hydrilla verticillata* (L.f.) Royle, were collected from various aquatic habitats of the district Bhimber, Azad Jammu and Kashmir. The plants were preserved in the fixative solution. Then, they were subjected to the anatomical analyses.

In the stem sections of *Nymphaea alba* L., when examined under the light microscope (40X), a layer of epidermis was clearly observed surrounded by Trichomes. The Trichomes were profusely

surrounding the layer of the epidermis. The length of the Trichomes was observed as  $4.50 \pm 0.77$  ( $\mu\text{m}$ ) ranging from  $3.85\mu\text{m}$  to  $5.36\mu\text{m}$ . However, the width of the Trichomes was observed as  $2.11 \pm 0.48$  ( $\mu\text{m}$ ) ranging from  $1.56\mu\text{m}$  to  $2.48\mu\text{m}$ . Hanif *et al.* 2017 reported the similar anatomical features in *Eichhornia crassipes* (Mart.) Solms and *Nymphaea rubra*. While studying the *Nymphaea alba* L., aerenchyma were clearly observed, significantly indicating the anatomical adaptation in their aquatic habitats in response of excessive water supply. The length of the aerenchyma was observed as  $121.50 \pm 47.34$  ( $\mu\text{m}$ ) ranging from  $71.03\mu\text{m}$  to  $164.94\mu\text{m}$  in different aerenchyma. However, the width of the aerenchyma was observed as  $92.76 \pm 32.26$  ( $\mu\text{m}$ ) ranging from  $70.43\mu\text{m}$  to  $129.76\mu\text{m}$  in different aerenchyma in of the stem of *Nymphaea alba* L. Hanif *et al.* 2017 reported the similar results and suggested the presence of aerenchyma as a main anatomical feature of the hydrophytes to adapt to their ecological habitat. They suggested the presence of six larger aerenchyma surrounding the inner vascular bundle. Jung *et al.* (2008) also identified the anatomical pattern of aerenchyma in hydrophytes.

Xylem and Phloem were clearly observed in the inner vascular bundle. The length of the xylem was measured as  $17.91 \pm 1.62$  ( $\mu\text{m}$ ) ranging from  $16.32\mu\text{m}$  to  $19.56\mu\text{m}$ . However, the width of the xylem was measured as  $6.55 \pm 0.79$  ( $\mu\text{m}$ ) ranging from  $6.04\mu\text{m}$  to  $7.41\mu\text{m}$  in different samples of the stem of *Nymphaea alba* L. The phloem was larger in length and broader in the width than xylem. The length of the phloem was measured as  $29.57 \pm 1.97$  ( $\mu\text{m}$ ) ranging from  $27.62\mu\text{m}$  to  $31.56\mu\text{m}$ . However, the width of the phloem was observed as  $31.47 \pm 2.54$  ( $\mu\text{m}$ ) ranging from  $29.07\mu\text{m}$  to  $34.14\mu\text{m}$  in the phloem of different sections of the stem of *Nymphaea alba* L. Hanif *et al.* (2017) observed the poorly developed vascular bundles and mainly larger

aerenchyma were observed to adapt to the influx of excessive water in the ponds and aquatic habitats.

In the transverse sections of the stem of *Hydrilla verticillata* (L.f.) Royle, a compact arrangement of the cells was observed. Different cells and tissues such as collenchyma cells, epidermal cells, ground tissues, xylem, and phloem were present and observed using the light microscope (40X). The existence of different sized lacuna, collenchyma cells, parenchyma cells, multicellular trichomes, and distribution of chloroplast in various regions of the stem of hydrophytes were also reported by Jayeola and Folorunso (2009).

The transverse section of the stem of *Eichhornia crassipes* (Mart.) Solms was observed under the light microscope (40X). Aerenchyma cells were prominently observed. Vascular bundles were present in which xylem and phloem were observed. The length of the aerenchyma cells was measured as  $40.50 \pm 1.00$  ( $\mu\text{m}$ ) ranging from  $39.58 \mu\text{m}$  to  $41.57 \mu\text{m}$ . However, the width of the aerenchyma cells was observed as  $28.25 \pm 1.48$  ( $\mu\text{m}$ ) ranging from  $26.56 \mu\text{m}$  to  $29.35 \mu\text{m}$ . Similar findings were also observed by Hanif *et al.* (2017) who studied the adaptations of hydrophytes. Larger aerenchyma were reported with the abundant trichome density. Multicellular trichomes were observed. The selected hydrophytes were belonging to different families. However, it was pragmatically observed that, all these hydrophytes were significantly similar in their anatomical features and characteristics to adapt to their ecological environment (i.e. aquatic habitat). These anatomical characteristics are commonly found among aquatic angiosperms and represent and represent adaptations in the aquatic habitats. It was also investigated that similarity in the anatomical features of selected plants of a geographical locality, although belong to different families, is related to the

similarity of their habitat (i.e. aquatic habitat) also described by Hanif *et al.* (2017).

This study involves the anatomical characterization of various structures in the transverse sections of the stem of some selected hydrophytes, collected from district Bhimber, Azad Jammu and Kashmir. The anatomical characteristics of these hydrophytes were not investigated before by using microscopic techniques. This thesis provides the evident micrographs of the selected plants along with the measurement of different cells examined in the stem of these plants which will be a significant addition to be studied and utilized as the foundational data base in the regional scientific literature and worldwide.

## CONCLUSION

In the present research, anatomical characteristics of the transverse sections of the stem is investigated to comprehend their adaptations to their ecological environment. Larger aerenchyma and multicellular trichomes were observed, indicating that these species (i.e. *Eichhornia crassipes* (Mart.) Solms, *Hydrilla verticillata* (L.f.) Royle, and *Nymphaea alba* L.) were better adapted to their aquatic environment.

## FUNDING STATEMENT

The authors declared that no funds, grants, or other support were received during the conduction of study and preparation of this manuscript”.

## AUTHOR’S CONTRIBUTION

UH and MUF contributed to the study conception and design. Material preparation, sample and data collection and analyses were performed by RA. The first draft of manuscript was written by RA, and all authors commented on previous versions of manuscript. The final draft was written by WA. All authors read and approved the final manuscript.

## DATA AVAILABILITY STATEMENT



All data presented is primary and included in the manuscript

## ACKNOWLEDGMENTS

Thanks to the Department of Botany, Government College University, Lahore, for providing research facilities.

## CONFLICT OF INTEREST

Authors have no relevant financial or non-financial conflicts of interest to disclose.

## REFERENCES

- Ajaib, M., Z. U. D. Khan, M. F. Siddiqui. 2012. Ethnobotanical study of useful climbers/twiners of district Kotli, Azad Jammu and Kashmir. *International Journal of Biology and Biotechnology*, 9(4): 421-427.
- Ayanda, O.I., A. Tolulope, P. Femi. 2020. *Eichhornia crassipes* (Mart.) Solms: uses, challenges, threats, and prospects, *The Scientific World Journal*, 2020: 3452172.
- Bavaru, A. B. Rodica. 2002. Morfologia și anatomia plantelor, Ed. ExPonto
- Bergholt, N. L., H. Lysdahl, M. Lind, C. B. Foldager. 2019. A standardized method of applying toluidine blue metachromatic staining for assessment of chondrogenesis. *Cartilage*, 10(3): 370-374.
- Bomblies, K., V. Shukla, C. Grajam. 2008. Scanning Electron Microscopy (SEM) of plant tissues. *Cold Spring Harbor Protocols*. 3(4): 1-3.
- Coetzee, J. A., M. P. Hill, D. Schlange. 2009. Potential spread of the invasive plant *Hydrilla verticillata* in South Africa based on anthropogenic spread and climate suitability. *Biological Investigations*, 11: 801-812.
- Hanif, U., M. Khalid, S. Ishtiaq, S. Shaheen, T. Cheema and A. K. Achakzai. (2017). Assessment of morphoanatomical study of *Eichhornia crassipes* (Mart) Solms. And *Nymphaea rubra* Roxb. Ex. Andrews. A Step toward minimizing the adulterations in drug plants. *Transylv. Rev.*, 25: 4737-4744.
- Harrison, H. Y. 1955. *Nymphaea* L. *Journal of Ecology*, 43(2): 719-734.
- Jafari, N. 2010. Ecological and socio-economic utilization of water hyacinth (*Eichhornia crassipes* Mart Solms). *Journal of Applied Science and Environmental Management*, 14: 43-49
- Jain, S. K., R. R. Rao. 1977. A Handbook of Field and Herbarium Methods. Today and Tomorrow Printers and Publishers, New Delhi.
- Jayeola, A. A., E. A. Folorunso. 2009. Ecological anatomy of some hydrophytes in Nigeria. *African Journal of Biotechnology*, 8(14):1-4.
- Jones, J. I., W. Li, S. C. Marberly. 2003. Area, altitude and aquatic plant diversity. *Ecography*, 26(4): 411-420.
- Jung, J., S. C. Lee, H. K. Choi. 2008. Anatomical patterns of aerenchyma in aquatic and wetland plants. *Journal of Plant Biology*, 51: 428-439.
- Langeland, K. A. 1996. *Hydrilla verticillata* (L.F.) Royle (Hydrocharitaceae), "the perfect weed." *Castanea*, 61: 293-304.
- Lux, A., S. Morita, J. Abe, K. Ito. 2005. An improved method for clearing and staining free-hand sections and whole-mount samples. *Annals of Botany*, 96(6): 989-996.
- Maqbool, M., M. Ajaib, B. K. Hayat, M. Ishtiaq, K. Humaira, H. Tanveer. Traditional knowledge based inventory of wild plants of Watala National Park and allied villages from Bhimber District, Azad Jammu and Kashmir, Pakistan. *Applied Ecology and Environmental Research*, 17(5): 12023-12055.
- Michel, A., R. S. Arias, B. E. Scheffler, S. O. Duke, M. Netherland, F. E. Dayan. 2004. Somatic mutation-mediated evolution of herbicide resistance in the nonindigenous invasive plant (*Hydrilla verticillata*). *Molecular Ecology*, 13: 3229-3237.
- Milne, M. J., K. Kearins, S. Walton. 2006. Creating adventures in Wonderland: The journey metaphor and environmental sustainability. *Organization*, 13(6): 801-839.
- Pieterse, A. H. 1981. *Hydrilla verticillata*—a review. Abstracts on tropical agriculture. Vol. 7, Royal Tropical Institute, Amsterdam, pp. 9-34.

- Quesnel, L. B. 1971. Chapter I Microscopy and Micrometry. In *Methods in Microbiology*; Norris, J.R., Ribbons, D.W., Eds.; Academic Press: Cambridge, MA, USA, 1971; Volume 5, Part A, pp. 1–103., University of Edinburgh.
- Rodrigo, M A. 2021. Wetland restoration with hydrophytes: A Review. *Plants*, 10(6):1035.
- Ronzhina, D.A. and V.I. P'yankov. 2001. Structure of the photosynthetic apparatus in leaves of freshwater hydrophytes: 2. Quantitative characterization of leaf mesophyll and the functional activity of leaves with different degrees of submersion. *Russian Journal of Plant Physiology*, 48: 723-732.
- Ruzin, S. E. 1999. Plant microtechnique and microscopy. Oxford University Press, New York.
- Steward, K. K. 1993. Seed production in monoecious and dioecious populations of Hydrilla. *Aquatic Botany*, 46: 169-183.
- Sumithran, S., S. Raj, P. J. Sanjeeva. 2013. Keystone functions of *Hydrilla verticillata*. Department of Biological Sciences, Eastern Kentucky University Richmond, USA.
- Tiner, R.W. 2006. Lists of potential hydrophytes for the United States: a regional review and their use in wetland identification. *Wetlands*, 26(2): 624-634.
- Verkleij, J. A. C., A. H. Pieterse, G. J. T. Horneman and M. Torenbeek. 1983. A comparative study of the morphology and isoenzyme patterns of *Hydrilla verticillata* (L.f.) Royle. *Aquatic Botany*. 17: 43-59.
- Werker, E. 2000. Trichome diversity and development. *Annual Review of Plant Biology*, 51\*(1): 645-675.
- Wheeler. B. D. 1997, *Aquatic Plants in Britain and Ireland*. By C. D. PRESTON and J. M. CROFT. 137: 371-372.
- Xia, H., X. Ma. 2006. Phytoremediation of ethion by water hyacinth (*Eichhornia crassipes*) from water. *Bioresource Technology*, 97: 1050–1054.
- Yeo, R. R., R. H. Faulk, J. R. Thurston. 1984. The morphology of hydrilla [*Hydrilla verticillata* (L.f.) Royle]. *Journal of Aquatic Plant Management*, 22: 1-17
- Zelko, I., A. Lux, T. Sterckeman, M. Martinka, K. Kollárová, D. Lišková. 2012. An easy method for cutting and fluorescent staining of thin roots. *Annals of Botany*, 110(2): 475–478.
- Zhao, Q., J. Bai, L. Huang, B. Gu, Q. Lu. 2016. A review of methodologies and success indicators for coastal wetland restoration. *Ecological Indices*. 60, 442–452.