## REVIEW ON MORPHOMETRICS AND MANAGEMENT OF BACTROCERA ZONATA (DIPTERA: TEPHRITIDAE), AN ECONOMIC PEST IN PAKISTAN

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#### Abstract

Bactrocera zonata (Saunders), Diptera: Tephritidae, is a voracious pest. Many host plants are attacked by B. zonata which causes a loss of \$200 million per year in Pakistan. The hosts of B. zonata are guava, mango, peach, apricot, citrus, and fig. Climate change affects the metabolic processes of insects as an increase in temperature delays the process of insect development, and affects their survival rate, and longevity. Temperature affects the growth stages and development of B. zonata from larval emergence to adulthood. The behavioral responses shown by insects are governed by volatile compounds released by insects and host plants. Factors affecting host plant selection involve host finding and host acceptance. Host finding and host acceptance are controlled by vision, mechanoreception, olfaction, and contact chemoreception. Exogenous and endogenous factors influence the activity of insects. The intensity of light and temperature can limit the locomotory, flight, and foraging behaviors of fruit flies. B. zonata is a destructive pest and difficult to control due to its ecological, and morphological behaviour. Management of fruit flies is difficult. Several control tactics, including wrapping, covering, and spraying artificial pesticides are being used. For immediate control, farmers use insecticides such as diptrex, triazophos, and imidacloprid. The excessive use of pesticides has led to many issues which made scientists believe in biological control measures. The best practice to control any pest is IPM, which can replace other practices as well, as agrochemicals Entomopathogens, predators, parasitoids, and bait traps of botanicals are used worldwide for biological control.

Keywords: Behavior, Climate Change, Fruit Fly, IPM, Mechanoreceptions, Predators

## **Introduction:**

B. zonata is native to tropical Asia and widely distributed in Indonesia, Nepal, Oman, India, Bhutan, and Pakistan (Delrio and Cocco, 2012). B. zonata is a polyphagous and quarantine pest with a wide host range, causing a loss of \$200 million in fruit exports per year (Delrio and Cocco, 2012). Fruit flies are distributed along the coastal, subcoastal, semi-desert, and northern plains of Pakistan. In 2017, the species was reported in the Sargodha district from April to May, with the highest populations recorded in February and October. In climate change scenarios, pests can shift their host range according to environmental

conditions (Arif *et al.*, 2017). The distribution of *Bactrocera* spp. in Pakistan is presented in Table 1.

It was also observed in some arid regions and Punjab's levellands. Because it is a rare pest, the hilly regions of Peshawar and Islamabad were also alerted (Marwat *et al.*, 2013). Although *B. zonata* is active all year round, March, May, July, and August were the months with the highest seasonal abundance. The pest breeds in the spring, overwinters as a pupa, and hibernates during the winter (Kapoor, 1986). Fruit flies are found all throughout the world, disrupting international trade and incurring enormous financial losses to fruits and vegetables.

#### **Host Plants:**

*B. zonata* is a polyphagous pest with the ability to fly 25 miles or more in search of a host plant. There are approximately 40 species of fruit that are attacked by *B. zonata*. The hosts of *B. zonata* are guava, mango, peach, apricot, citrus, and fig.

## Biology and ecology:

Bactrocera zonata (Saunders) Tephritidae: Diptera have four growth stages: eggs, larvae, pupae, and adults. The first stage of *B. zonata* development is an egg laid by a female under the skin of the fruit. The eggs are creamy white, elongated, and elliptical in shape, approximately 1.0 to 1.2 mm long, with a distinct micropyle. Eggs hatch into larvae within 3–30 days at 35–15°C (Binyameen *et al.*, 2021). The entire life cycle is

completed within 15 days at 35 °C and 60 days at 15 °C (Adly, 2016).

The larvae consisted of a rolled spiral head, three thoracic segments, and eight abdominal segments (Balmès and Mouttet, 2017). Mouth hooks, anterior spiracles, and posterior spiracles are special characteristics of B. zonata that change during larval development (Murtaza et al., 2021). B. zonata undergoes three larval instars. The respiratory system of *B. zonata* consists of two pairs of spiracular openings at the prothorax and posterior side of the abdomen. The first larval instar is 0.07 to 0.09 in length and white in color. The posterior end of larvae is broad and round, whereas the anterior end is tapered (Balmès and Mouttet, 2017). Yellowish-brown mouth hooks are present on the head. The body is semi-transparent, and the skeleton is visible near the head region (Fadul, 2021).

Table 1: Distribution of Bactrocera spp. in Pakistan

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Province	Species	Reference
Punjab	B. dorsalis, B. zonata, and B. cucurbitae	(Ahmad et al., 2019)
	B. dorsalis, B. zonta, B. cucurbitae, and B. correcta	(Noman et al., 2021)
	B. zonata	(Khan and Naveed, 2017)
Sindh	B. dorsalis, and B. zonata	(Zain-Ul-Aabdin Abro et al., 2020)
Balochistan	B. zonata	(Khan et al., 2016)
	B. zonata, B. tau, B. invadens,	(Maula et al., 2022)
	B. cucurbitae, and B. dorsalis	
	B. cucurbitae	(Qureshi et al., 2017)
Khyber Pakhtunkhwa	B. dorsalis, B. correcta, B. cucurbitae, B. signata,	(Kausar et al., 2022)
	B. scutellaris, B. tau, B. nigrofemoralis, B.	
	zonata, and B. zahadi	

B. zonata lays white to creamy yellowish eggs under the fruit peel. One female lays a maximum of 93 eggs per day, and approximately 564 eggs are laid by the females throughout their

lifetime (Awad, 2012The adult emerges from the pupae and sexually matures within 2-3 weeks at 35-15°C. Females can live for 82-112 days.

Environmental factors influencing the growth and development of *B. zonata*. An optimum

temperature of 25°C to 35°C and humidity of 40-60% is necessary for *B. zonata* survival (Khan and Naveed, 2017). The largest population of *B. zonata* was seen in June, July, and August, while the following months saw a fall in population (Khan and

Naveed, 2017; Murtaza *et al.*, 2021). Growers in Pakistan face problems with fruit and vegetable infestations. Management of *B. zonata* can be effectively implemented with proper knowledge of its life cycle.

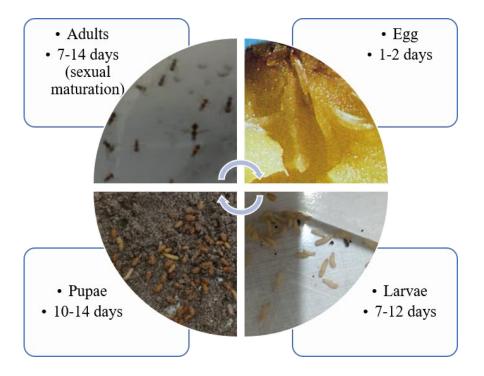


Figure 1: Life cycle of Bactrocera zonata

Bactrocera zonata attacks peaches, guava, mangoes, papaya, and citrus. It can fly for approximately 15 miles in search of a host plant. Bactrocera zonata competes with other species of Tephritidae fruit flies due to its short developmental phase, high reproductive rate, and survival. Compared to its competitors, B. zonata can endure lower relative humidity and can survive buried in soil for a longer period than its pupae. Adult fruit flies feed on nectar, rotting fruits, and plant sap for development and reproduction. The feeding behavior of B. zonata was temperature-dependent, and the most efficient feeding behavior was observed at night. Under field conditions, the fruit fly completed development at 10°C. The ideal temperature for the development of feeding, egglaying, and pupation was observed to be 25 to 29°C. The fruit flies remained active from early March to mid-November. After 10 to 16 days of adult emergence, oviposition occurs in the fruit skin with a 38.1 mm diameter.

## Oviposition behavior of *B. zonata*:

B. zonata shows two types of oviposition characteristics: pre-alighting and post-alighting behavior. These behaviors have a crucial role in population growth. It was revealed that the maximum number of pupae and adults were formed in guava (Murtaza et al., 2021). Analysis of volatile chemicals present in the fruit showed that some aromatic and aliphatic esters are the major factors for egg-laying B. zonata. Guava volatiles play an important role in the attraction and oviposition of B.

zonata (Binyameen et al., 2021). Another experiment was conducted to determine B. zonata's oviposition preference for guava, banana, chikoo, ber, apple, and citrus. In this case, guava is preferred (Rauf et al., 2013). Species-specific competition between the Tephritid species, Anastrepha fraterculus, and Ceratitis capitata, was used to evaluate the species interaction. They rejected the fruits, which were previously infested by conspecific and heterospecific individuals. The results suggest that the infestation of Tephritid species may vary due to the recognition of heterospecifics. Cross-recognition is a beneficial trait in nature due to which inter-specific competition is reduced and results in the coexistence of species (Liendo et al., 2020).

Adnan *et al.* (2020) experimented on sexual compatibility, and mating performance was studied under semi-field conditions. It has been reported that the average mating time was observed at 5-6 hours for *B. zonata*. Ndzana Abanda *et al.* (2016) evaluated the sexual competitiveness of males in SIT programs. Attraction to methyl eugenol is an age-dependent factor that accelerates the sexual maturity of *B. zonata*. showed some behavioral effects when exposing its antennae to some volatile compounds. Pyrazines, ethyl esters, and spiroacetals were identified using SSGA. These volatile chemicals are released by fruit flies and increase in quantity, specifically with age (Levi-Zada *et al.*, 2020).

Fruit flies also show an attraction to mango extract, as they have some electro-physiological responses to mangoes (Aluja, 2020). Approximately 14 species of host plants are preferred by fruit flies. The most preferred hosts for fruit flies are mangoes and bananas. Fruit flies also exhibit color preferences and behavioral responses. Trap color preference in the field was evaluated, and the results showed that trap color affected the preference of

fruit flies. Yellow traps are the most preferred for fruit flies. Some behavioral reactions are affected by trap colors, such as host finding, flower visitation, egg deposition, and proboscis extension. This study shows that there are some photoreceptor sensitivity characteristics in flies. Fruit flies can imagine colors and differentiate between them (Lunau, 2014).

The mating and courtship behaviors of phytophagous insects depend on the host plant. Females simply visit the host plant for oviposition and feeding purposes, while males search for host plants for females. Oviposition punctures of *B. zonata* result in the invasion of saprophytic microbes, which results in the rotting of the fleshy tissues of the fruit. After the emergence of larvae, they start feeding on flesh, which results in fruit decay. Infested fruits fall to the ground, which facilitates the pupation of *B. zonata*. On the other hand, it also causes economic loss by reducing mango exports (Myers *et al.*, 2016).

## Environmental factors affecting *B. zonata*:

Temperature and humidity, for example, have an important impact on the population growth of fruit flies. July and June were the months of greatest population growth for B. zonata (Noman et al., 2021). Using maximum and minimum temperatures, rainfall, and humidity, evaluated the growth and development of B. zonata (Khan et al., 2021). The maximum population of *B. zonata* was observed at peak temperature when the mango fruits were ripped. Rainfall and humidity were negatively correlated with the development of B. zonata. (Chiu et al., 2019) studied the temperature influence on transgenic and natural B. dorsalis development. The population was exposed to 30°C, 32.5°C, and 35°C, The growth and development of both fly strains were negatively affected. High temperatures affected life history traits (Ndlela et al., 2021) Temperature affects the growth stages and

development of *B. zonata* from larval emergence to

adulthood. The geographic pattern of fruit fly occurrence and their behavior were studied under laboratory conditions by managing at different temperatures. From 15 to 30°C, the development process decreased as the required temperature increased. There was a significant difference in temperature between the four incubation periods. Egg development time was significantly reduced as temperature decreased from 15 to 25°C, not between 25 and 30°C. Similar to the temperature range of 15 to 30°C, the development process of larvae and pupae decreases as temperature increases. The larval and pupal lower thresholds for the development were 13 and 10.9°C. The population rate was highest in B. zonata at  $87 \pm .48$  at 25°C, which is considered the optimal temperature for growth. Based on these results, fruit fly population dynamics can be predicted, which will help make management tools.

Spatio-temporal variations of *B. zonata* were studied in the context of temperature fluctuations. According to risk indices, 59.69% of the area is highly suitable for *B. zonata* in India (Choudhary *et al.*, 2021). The population growth model predicts that the optimum temperature for survival of the *B. zonata* population is in the range of 25 to 30 Results reported that the intrinsic rate and finite rate were maximum at 30°C while at low temperatures they were in the range of 15 The population was unable to lay eggs. Hence, temperature shows prime importance in population growth (Choudhary *et al.*, 2020).

As population growth is highly temperature-dependent, diets given to immature *B. zonata* also play an important role. Ben-Yosef *et al.* (2021) investigated the nutritional importance of larval diets by manipulating the diet by adding yeast at 2 to 10 percent and evaluating the heat and cold tolerance by population. A yeast-deficient diet resulted in significant adult weight loss. Yeast-rich diets showed a significant tolerance level for heat

stress. There was no significant difference in adult survival based on larval diet composition. Acute cold stress (3–3°C for 2 hours) did not have a significant effect on adult survival. Nutritional heat-tolerance mechanisms regarding diets were discussed.

In the climate change scenario, temperature extremes play a major role in increasing pest populations. Ullah *et al.* (2022) reported that temperature extremes at low and high levels significantly increase larval development, pre-adult durations, APOP, TPOP, and mean longevity of *B. zonata*. The findings of the experiment showed that the stress of temperature increased larval development, reproduction rate, and life span of *B. zonata*. Life history traits that remain under the influence of temperature might be changed and become a key strategy for pest management.

#### Interspecific Competition

He et al. (2023) witnessed the phenomenon of interbreed competition in insects, notably among akin species, which is frequently concomitant with an encroachment process. Tephritid incursions in regional populations brought about alterations in interspecific populace contests, culminating in the eradication of local colonies. (Courtney and Kibota, 2017). The comprehension of such rivalry in Tephritidae could be conceptually elucidated by the inherent mechanisms and validate the hierarchical structure and biological assessment markers that are employed for forecasting the capacity and achievement of invasive Tephritid fruit flies (Charlery De La Masseliere et al., 2017).

The egg-laying inclination of both *B. correcta* and *B. dorsalis* towards various hosts displayed significant selectivity; hosts with a favorable aroma were highly favored for the growth and maturation of their offspring. Relying on a sole parameter (survival rate, pupal weight, or development period) was insufficient to assess the host adaptation of

tephritids; it was discovered that there was a convergence between both species differentiation in ecological habitats. the larval stage, there was a notable occurrence of elevated population density and intense competition. Nevertheless, B. correcta larvae exhibited greater resilience in such crowded conditions compared to B. dorsalis, elucidating their competitive prowess. The mating behavior of both species displayed dissimilarities, revealing a negative influence on each other in an uneven manner. Three species, B. correcta, B. dorsalis, and Ceratitis capitata, were also investigated concerning their interactions, which revealed that there is no interbreeding among them. However, oviposition competition was evident in guava at the optimal temperature, and the two Bactrocera species had a significant inhibitory effect on C. capitata (He et al., 2023).

In the majority of Dacines, there is a form of "scrambles" competition among larvae, leading to a certain level of pressure, causing the resulting pupae and adults to reduce in size as larval density per fruit rises. Diminutive females exhibit lower fecundity, possess a reduced number of ovarioles, and face a higher likelihood of survival challenges, especially considering Australia's suboptimal weather conditions (Inskeep, 2019). Despite multiple sympatric dacines breeding in the same fruit, there is scarce evidence of interspecific conflicts among them (Castro-Vargas et al., 2023). Senior larvae inflict significant damage on younger larvae within the same fruit through an unidentified mechanism; competition for resources occurs among larvae of the identical age group (Kay, 2023).

While *B. dorsalis* was limited, *B. zonata* made up to 98% of all males collected in India using GK hooks baited with methyl eugenol, according to studies (Baker *et al.*, 2019). *Anastrepha striata* adults attacked and pushed away *C. capitata* adults

from guava fruits, based on findings reported by Paranhos *et al.* (2021). While *A. obliqua* and *C. capitata* were found present in 94% of mangoinfested fruits (Petitinga *et al.*, 2021) and 6% of them, respectively.

In 60.4% of relationships, A. obliqua in Central America scared *C. capitata* off mangoes by attacking. The occurrence of competitive conduct was not present in 35.8% of the occurrences. A. *obliqua* appeared to be evicted from the fruits by C. capitata in the remaining 3.8% of observations Ghanim, 2009, A. obliqua prefers mango fruits as a landing spot, but C. capitata prefers oranges. According to interspecific competition study, A. obliqua is a stronger opponent than C. capitata. A. obliqua's reaction to the presence of C. capitata is to first circle the smaller fly before charging directly at it and driving it off the fruit, the difference in size could have a role. The fact that A. obliqua is three times bigger than C. capitata may be the cause of this competitive superiority. Similarly, abundance of fruit flies is normally influenced by fruit. In intraspecific competition, among B. zonata and C. capitata populations, B. zonata was the predominant population due to its dominance and environmental potential (Ghanim, 2009).

Duyck *et al.* (2022) examined interspecific competition mechanisms among polyphagous fruit flies (Diptera: Tephritidae) species. In both larval and adult interfering rivalry, these species displayed an unbalanced and hierarchical interaction among themselves. During the three consecutive seasons, *B. zonata* replaced *C. capitata* at a higher rate on eight distinct host plants, including mandarin, navel, mango, Valencia oranges, balady, apricot, guava, and peach. (Abd-Elgawad, 2021). The most dominant population from March to May in different orchards was *B. zonata* as compared to the Mediterranean fly (Boulahia-Kheder, 2021b). The presence of *Bactrocera invadens* (Drew) in 350

people was associated with issues with *Ceratitis cosyra* and other closely related fruit fly species (Ali *et al.*, 2014). Interspecific competition appears to be the cause of this behavior, which could account for the observation of *B. invaders* while *C. cosyra* dominated emergence from the hatched fruits of alternate host plants by as much as 87%.

For oviposition, females spend some time on host selection. The better the host, the greater the oviposition, larval development, and survival. The chemical cues released by host plants determine the tephritidae host selection range. Polyphagous insect species respond more strongly to visual messages and plant-released volatile compounds (Aluja, 2020). The feeding, fecundity, and growth habits of insects may be directly affected by plant metabolites and nitrogen content (Jaworski and Hilszczański, 2013).

The fruit fly *B. tryoni* showed preferences for oviposition in citrus fruits. Compared to the lemon and other cultivars, the grapefruit was discovered to be the most favored fruit for oviposition (Muthuthantri and Clarke, 2012). Similar findings were also demonstrated by Prakash and Kaur (2022), which determined the ovipositional preferences of *B. zonata* in various fruits such as apples, guavas, citrus, and berries.

# Behavioral reactions of *B. zonata* adults in response to some olfactory stimulants:

The behavioral responses shown by insects are governed by volatile compounds released by both insects and host plants. The insect response toward attractant pheromones is elicited by the various semiochemicals of the host plant (Quilici *et al.*, 2014). In many of the findings, it was suggested that peach fruit flies prefer to lay eggs in a suitable host. Their ovipositional preferences are influenced by many factors, such as the topography of the area, climatic conditions, and fruit odors, sizes, and colors (Lux *et al.*, 2017). In another finding, it was

reported that fruit color, size, odor, skin type, and foliage influence host insect selection preferences (Salman *et al.*, 2022).

Methyl eugenol is the most effective allure for male flies of *Bactrocera* spp. and *Ceratitis* species, excluding those belonging to the subgenus *B. zeugodacus*. Some flies of the Dacine subgenus showed a response to methyl eugenol and cure lure. The effective range of methyl eugenol at very low concentrations is u to 1 km (Manrakhan 2016; Manrakhan *et al.* 2017; Manoukis *et al.* 2017 Yesmin 2019). Allelochemicals are generated through ingestion for specific functions like defense and sexual communication (de Souza *et al.*, 2022).

This unique phenomenon takes place in the interaction of insects with plant materials, including phenylpropanoids and Tephritid fruit flies. For defensive purposes, the insects assimilate the plant chemicals directly or convert them into derivatives through feeding on plant tissues. Methyl eugenol proved to be effective when combined with various insecticides (like malathion, dichlorvos, or toxic baits) to detect and monitor male *B. zonata* flies in orchards (Navarro-Llopis and Vacas, 2014). (Drew (1987) and Hosseini *et al.* (2017) proposed that the fruiting host plant functions as a pheromone attracting males and a food attractant for females. Primarily, *dacine spp* react more strongly to olfactory stimuli than to visual ones.

As a result, trapping using male lures (Trimedlure and methyl eugenol) is essential for quarantine surveys and fruit fly specimen identification. Ghanim (2009) highlighted that *B. zonata* exhibited a response to methyl eugenol and combinations of Dorslaure, as well as mixtures of Trimedlure and Capilure in the field. The baited traps with malathion and methyl eugenol were used to investigate seasonal fluctuations of *B. zonata* and *B. dorsalis* Bagheri *et al.* (2017). In Pakistan, *B.* 

zonata can be monitored using baited traps with methyl eugenol, male lure, and insecticides. Traps containing cotton wicks treated with methyl eugenol were effective for 9 weeks duration. (Draz, 2016).

Methyl eugenol along with plant-derived attractants is used for the management of *B. zonata* (Amin, 2017). In Pakistan, an extract from *Ocimum sanctum* leaves was found to be a powerful attraction that could attract *B. zonata* from orchards up to 0.8 kilometers away. Ghanim (2009) documented that a trap containing a glass petri dish with five drops of methyl eugenol successfully trapped a total of 49.36 male fruit flies in a sapote orchard. The trapped species included *B. dorsalis*, *B. correcta* and *B. zonata*. Males of *Ceratitis*, *Dacus*, and *Bactrocera* can be collected using synthetic bait traps also known as para pheromones.

Male adults of B. zonata were caught using protein hydrolyzate as bait and the attractant methyl eugenol mixed with 50% EC Malation (Agarwal et al., 1999; Khosravi et al., 2018). B. zonata, methyl eugenol-baited traps loaded with three pesticides (Diptrex, Laser, and Methomyl) were placed in guava plantations in Pakistan at several elevations (1, 1.5, and 2 m). Diptrex emerged as the most effective insecticide, with the traps placed at a height of one meter proving to be the most successful. Mixture of yeast brewery waste which was prepared by local protein bait was used in comparison of imported bait through McPhail traps which were significantly effective in management of *B. zonata*. Gupta and Bhatia (2000) and Stanley et al. (2015) conducted population monitoring of B. zonata and B. dorsalis using bottle traps placed in guava and mango orchards. The highest capture of 98.6 and 517.0 males per trap for mixed populations was observed in mango and guava orchards. Ragab and Youssef (2021) assessed three attractants: an old local attractant (prepared one year ago), a 2%

solution of di-ammonium phosphate, and a fresh local attractant (comprising molasses, brewer's yeast, and water) at a concentration of 15%, for attracting *B. zonata* using McPhail traps in fig orchards located in the north-western coastal region. The capture-to-detection (CTD) ratio of *B. zonata* varied between 0.04 and 0.69, 0.02 and 1.49, and 0.02 and 0.07 flies when fresh local attractant, diammonium phosphate, and old local attractant were employed, respectively.

Anjum et al. (2000) evaluated the effectiveness of diffused methyl eugenol in controlling B. zonata and B. dorsalis in mango orchards. The 2 to 6 traps were hanged pheromone traps per hectare and concluded that four pheromone traps were effective control for these pests. The male annihilation method of managing B. zonata in orchards was tested using soaked-wood blocks as a substitute for the plastic traps now in use (Afzal et al. 2000). Each block was the best option for onfarm fly control in every situation, killing nearly twice as many flies and lasting nearly twice as long as a plastic trap. According to Amin (2013), the most effective method for delivering methyl eugenol was the cotton wick dispenser. Methyl eugenol was the most successful attractant among those tested, attracting the greatest number of *B. zonata*.

Hasnain *et al.* (2023) evaluated a number of substances for their capacity to draw *B. zonata* to guava orchards. Ammonium acetate, di-ammonium phosphate, ammonium sulfate, ammonium hydroxide, and ammonium chloride were the compounds that produced the following daily trapping rates: 30.52, 28.74, 25.58, 17.15, and 17.56 adults, respectively. Compared to males, females showed a stronger affinity to ammonium compounds. *B. zonata* was less attracted to buminal and egy-lure (food attractants) than to ammonium compounds.

Sidahmed *et al.* (2014) utilized wet wooden blocks loaded with methyl eugenol to create a device for *Bactrocera* spp. According to Afzal and Javed (2001), oblong plywood shapes might be the best shape and material for lethal blocks in male annihilation that target *Bactrocera* spp. fruit flies. Using McPhail traps on gradually mature fruits, Amin (2013) evaluated the effectiveness of three distinct concentrations of di-ammonium phosphate (1%, 2%, and 3%), in attracting *B. zonata* during a one-year period. There were three concentrations for which the weekly mean of captured flies was ranked: 1% (59.82 flies/trap), 2% (62.38 flies/trap), and 3% (73.82 flies/trap). The concentration of 3% turned out to be by far the most appealing.

Buminal 5% (16.27 flies/trap), eugenol (3.39 flies/trap), di-ammonium phosphate 3% (40.11 flies/trap), and buminal 10% (13.25 flies/trap) were the other attractants that produced the weekly mean of captured flies, in decreasing order. All captured flies attracted by methyl eugenol and eugenol were male, whereas the remaining attractants captured both males and females. Arya et al. (2022) Cue-lure and methyl eugenol were tested for their efficiency against fruit flies B. cucurbitae and B. zonata infesting Momordica charantia. During the twenty-third standard week, the largest population peak was 395.6 and 297.3 per trap with cue-lure and methyl eugenol, respectively, whereas during the twentieth standard week, the top population was 423.3 and 396.3, respectively. Population dynamics and maximum temperature had a substantial positive association.

Using stiner-type traps at a consistent location, the population of *Bactrocera* fruit flies was monitored with two types of lure (cue-lure and methyl eugenol). *B. zonata* showed attraction to methyl eugenol from April to July. Moreover, Hee and Tan (2006) and Mandanayake *et al.* (2023)

Hemolymph has a role in transporting the attractantderived pheromonal components from the crop to the rectal gland organ, where the chemicals are released towards twilight, as demonstrated by the discovery of these pheromonal elements in the hemolymph of male *B. dorsalis* fed methyl eugenol. In order to draw adult B. zonata to citrus orchards, Saafan (2005) and Darwish (2014) evaluated a number of attractants, including bactrogel 1.3%, diammonium phosphate 2% and 3%, and buminal 5% and 10%. For buminal 5%, buminal 10%, diammonium phosphate 3%, and bactrogel 1.3%, the mean catch per trap per day (CTD) in the first season (2002/2003) was 0.14, 0.08, 0.05, and 0.06 flies, respectively. Females were more attracted to all tested chemicals than males. The traps (buminal 2.5%, buminal 5%, buminal 10%, di-ammonium phosphate 2%, and di-ammonium phosphate 3%) were effective in management of male and female B. zonata adults (Saafan, 2005b).

Elnagar *et al.* (2010) assessed three categories of attractants: sexual, olfactory, and food. Methyl eugenol lure demonstrated the highest trapping efficacy for *B. zonata*, specifically as a male attractant. Furthermore, food attractant lures showed the highest proportion of *B. zonata* females compared to olfactory traps. evaluated eight dispenser types (saturated with a blend of methyl eugenol and the toxicant DDVP).

## **Host preference:**

Factors affecting host plant selection involve host finding and host acceptance (Courtney and Kibota, 2017). Host finding and host acceptance are controlled by vision, mechanoreception, olfaction, and contact chemoreception (Bestea *et al.*, 2021). The biochemical and physical characteristics of fruits are linked to fruit ripening (Tangpao *et al.*, 2022). He *et al.* (2023) reported that the volatile chemicals of mango and guava play an important

role in fruit fly host preferences. The volatiles present in mangoes, which are associated with the ripening of fruit, are also involved in attracting fruit flies. Physical damage to fruit may facilitate the puncturing of fruit flies (Grechi *et al.*, 2021)

Infestation by fruit flies depends on the maturity stage of the fruit and the abundance of fruit flies (Drummond et al., 2019). Less than 4% fruit infestation was recorded in greenish, mature greenish, and early yellowish fruit stages. Maximum fruit invasion was recorded in ripe and overripe fruit stages, approximately 14-16% (Grechi et al., 2021). The peel texture of the fruit also facilitates the infestation of fruit flies. A comparative study of host preference revealed that the quality and quantity of the host affect the development of progeny. A maximum number of progeny was recorded in mangoes (Sarwar et al., 2013). Kumar et al. (2021) found significant variations in the growth, development, and net reproductive rate of fruit flies on four mango cultivars. The physical and biochemical characteristics of fruit play a role in the variation in growth and development of fruit flies.

Knowledge about host selection by fruit flies is crucial, as it helps the flies locate and assess hosts for oviposition. The flies use their sensory cues for food selection. B. zonata females lay eggs in delicate and tender portions of fruits by piercing through the ovipositor, and then larvae develop in the fruit. The peach fruit flies normally visit various fruits and then decide which host fruit is most preferable for oviposition (Joachim-Bravo et al., 2001). The female B. zonata locates the host distance and the source of food from a distance by using sensory receptors and odor receptors, respectively. After the host fruit selection on a morphological basis, the fruit flies land and take a taste bite to check the suitability of food for their progeny's development. The host fruit's rejection and acceptance are determined by the host fruit's

appearance, size, shape, and fragrance (Murtaza *et al.*, 2021)

Host utilization is not only dependent on the host fruit's nutritional content; it also depends on other factors. The hard fruit skin of unripe fruit is a limiting factor for fruit flies (Balagawi et al., 2005). The biological parameters of insects, such as larvae size, number of eggs, adult life, and ovipositional period length, are influenced by host fruit quality and type (de Oliveira et al., 2012). Fruit flies are attracted to fruits because of their lipid and protein content, which is required for egg deposition, locomotion, metabolism, flight, and sexual maturity. The water content is obtained through the environment, rain, or vapors (Koswanudin et al., 2018). The nervous system of insects is influenced by chemical cues that ultimately affect host selection. Fruit host susceptibility may alter its chemosensory responses. The banana and bitter gourd proved to be preferred hosts for B zonata and B. cucurbitae (Saeed et al., 2022)

Host fruit selection preferences depend on the landing states of fruit flies. In pre-and postlanding states, females reach the host fruit and then decide to oviposit there. When the fly encounters the host fruit, it first evaluates its suitability. The acceptance and rejection of the egg-laying site depend on the chemical and sensory responses of the host plant. The physical characteristics of the same plant change from species to species. The nutritional content, defense strategies, and physical states of host plants determine female flies' oviposition preferences (Balmès and Mouttet, 2017). In host preference, a suitable host's presence is crucial. About 40% of insects are host-specific. In the genus Bactrocera, various fruit flies are specific to hosts, while others are not. For oviposition, fruit flies select the vegetables by considering their progeny's performance (Salgado and Saastamoinen, 2019)

## Daily Rhythms of B. zonata:

The activity of insects is influenced by exogenous and endogenous factors. locomotory, flight and foraging behaviors of fruit flies are limited by the intensity of light and temperature (Gadenne et al., 2016). Bayoumy and Metwally (2017) evaluated the flight activity of *B*. zonata and Mediterranean fruit flies in guava orchards on an hourly basis. The male peach fruit flies took off before the females, but Mediterranean females and males took off at the same time. An increase in temperature above 30°C affects the flight of male rather than female flies. Temperature and male flight activities of fruit flies are inversely related, while a positive relationship between female flight activities and temperature was found. On the other hand, humidity is also negatively linked with male activity. A peach fruit fly male is attracted to sexual and olfactory attractants more than a Mediterranean fruit fly. Male catches were highest between 7:00 and 10:00 a.m., while female captures were highest between 11:00 and 5:00 p.m., reported that males fly actively in the morning while females fly actively in the evening in search of food and oviposition. This suggests that B. zonata and Mediterranean fruit flies are diurnal as well. Spray treatments for both species should thus be delivered between 7:00 a.m. and 10:00 a.m. for males and 11:00 a.m. and 5:00 p.m. for females.

Levi-Zada et al. (2020) have established behavioral impacts, or B. zonata antennae sensitivity, to pesticides. Through Solid microextraction and Gas chromatography, Mass Spectrometry analyses were performed to uncover volatiles unique to each sex of B. zonata that are released in a daily periodicity. The volatiles identified through chemical synthesis and GC-MS were pyrazines and ethyl esters. It was revealed that each volatile increase or decrease in number was specific to sex. Another technique that reveals the variation in the sensitivity of male and female

antennas to certain volatiles is Electroantennographic (EAG) dose response.

Bertolini et al. (2018) used Oxitec's selflimiting olive fly technology (OX3097D-Bol) to study the olive fly circadian clock based on evolutionary, genetic, anatomical, and behavioral responses which revealed that the circadian clock of olive fruit fly and Drosophila melanogaster are same. Changes in physiological conditions affect the expression of chemosensory receptors in *Bactrocera* dorsalis, which adapt to different states such as feeding, mating, and time of day. The study suggests that these receptors play a significant role in physiological processes related insect reproduction and survival Jin et al. (2017).

## Management of *B. zonata*:

B. zonata is a destructive pest and difficult to control due to its ecological, and morphological behaviour. Management of fruit flies is difficult. Several control tactics are used, including wrapping, covering, and spraying artificial pesticides (Stejskal et al., 2021). For immediate control, farmers use insecticides such as diptrex, triazophos, and imidacloprid (Dias et al., 2018). The excessive use of pesticides has led to many issues that scientists believe in biological control measures. Paranhos et al., (2019) reported that the excessive use of artificial termiticides resulted in resistance within four decades. The best practice to control any pest is IPM, which can replace other practices as well as agrochemicals (Deguine al., 2021). Entomopathogens, predators, parasitoids, and bait traps of botanicals are used worldwide for biological control.

## Physical control:

Sharma and Sanikommu, (2017) reported that bagging fruits and post-harvest quarantine treatments are the physical control methods. Fruit bagging keeps the fruit free from fruit fly attacks and

physical injury. This is undertaken just before the fruits reach the maturity stage when they are vulnerable to infection (Sarwar *et al.*, 2013). According to Amin *et al.* (2008), double-layer brown-colored bags are eco-friendly tools for managing fruit flies. The practice of fruit bagging should be adopted 40-55 days before the fruit setting.

#### **Cultural control:**

Cultural control practices include the implementation of tolerant variety, field sanitation practices, and eradication of infested fruits from the ground (Dias *et al.*, 2018). Manoukis *et al.* (2019) reported that cultural practices help to minimize *B. zonata*. *B. zonata* infests the mango peel and the larvae feed on the mango pulp due to which the mango quality is destroyed. Infested fruits must be buried 50 cm below the soil surface to minimize the infestation of *B. zonata*. Larvae of *B. zonata* can be destroyed by placing the infested fruits under the sun for 10 days (Manoukis *et al.*, 2019).

#### **Biological control:**

In comparison to pesticides, biological management is more affordable and less harmful to both people and the environment (Paranhos *et al.*, 2019). There are several methods of introducing biocontrol agents into pest control programs, including vaccination, inundation, traditional biological control, and conservation biological control. According to Dias *et al.* (2018) parasitoids, predators, natural enemies, and pathogens are frequently used in biological management, because they attack pests by eating them, parasitizing them, and spreading illnesses.

#### **Natural insect enemies:**

One method for reducing pest populations to tolerable proportions in conventional biological control programs is the deployment of exotic parasitoids and predators from the insect's native location (Adly, 2016). Parasitoids include *Psyttalia* 

sp., *Tetrastichus giffardianus*, and *Aganaspis* sp. Reported parasites include *Dirhinus giffardii*, a pupal parasitoid of *B. zonata* (Mahmoud *et al.*, 2019). The initial revival occurred approximately a month after the release date of the exotic parasitoid species. *Aganaspis daci* against *B. zonata*, as the incidence of parasitism reached 9.7%.

Ants interrupted them during oviposition and preyed upon mature Drosophila melanogaster, which was the main reason for the decline in fruit fly devastation (Mariano et al., 2019). The families Staphylinidae, Carabidae, Dermaptera, Chrysopidae, Formicidae, Coreidae, Coccinellidae, and Pentatomidae contain most fruit fly predators (Paranhos et al., 2019). Predatory mites, which also include insects, were evaluated when eaten on the eggs of B. zonata, including Amblyseius largoensis, Neoseiulus barkeri, Cydnosus negev, Typhlodromips swirskii, and Proprioseiopsis kadii (Momen et al., 2016)

## **Entomopathogens:**

Natural microbes called insect pathogens can be found in a variety of situations. At different stages of the insect host life cycle, these pathogens can cause sickness. It has been demonstrated that pathogens like nematodes, bacteria, and fungi are viable biological control methods for *B. zonata* (Shaurub, 2023)

Entomopathogenic nematodes cause their victims to die by introducing germs into the body (Aryal *et al.*, 2022). Some tephritid organisms have been observed to be severely affected by *Heterorhabditis* and *Steinernema* species. According to (Mahmood *et al.*, 2016), EPNs can infect *B. zonata* at various developmental stages. Nematodes can access dropping guava fruit larvae through the area where the soil surface and interface region meet.

They infect both pest adults and newly produced pupae in the soil as pest adults emerge

from their pupae. The newly discovered specimen, *Heterorhabditis marelatus*, demonstrated remarkable effectiveness against adults emerging from the pupal case of *B. zonata*, killing a significant portion of adults within 45 days after birth (Saleh *et al.*, 2019). Entomopathogenic nematodes have been used as biocontrol agents for many years. This may be improved by the isolation and selective breeding of more populations and species (Perry *et al.*, 2012). Parasites known as entomopathogenic fungi can prevent oviposition, decrease ovulation and reproduction of adult insect pests, and kill or paralyze them (Wei *et al.*, 2023).

According to (Valero-Jiménez *et al.*, 2016), the mode of action of EPF is initiated by the attachment of the spores and conidia to the host's cuticle layer, where they begin to germinate, grow, and eventually form a colony. *Metarhizium anisopliae* and *Baeuveria bassiana* are well-known EPF fungi (Ascomycota: Hypocreales) because of their distinctive traits and behaviors. They have several negative consequences on insects, such as impairment of molting, abrasion resistance, impairment of maturation, inhibition of oviposition, and significant mortality. Entomopathogenic fungi are among the most appealing biological control agents owing to these characteristics.

#### **Biopesticides:**

Biopesticides are eco-friendly and species-specific. Synthetic insecticides contain some chemicals which are harmful to non-target organisms. Biopesticides are not resistant to pests. Ugwu *et al*, (2021) experimented with the efficacy of three ethanol extracts of *Aframomum melegueta*, *Piper guineense*, *and Zingiber officinale* with a comparison of commercial botanical pesticides of AzaSol (6% azadirachtin) for the control of *B. dorsalis*. Pupation of *B. dorsalis* was targeted to evaluate the efficacy of ethanol extracts in the soil

of tree canopies. *P. guineense* and *A. melegueta* showed maximum control in comparison to AzaSol.

#### **Chemical Control:**

The most popular technique of control is chemical, and various pesticide formulations have shown success against different species of fruit flies (Vargas *et al.*, 2015; Dias *et al.*, 2018; Boulahia-Kheder, 2021; Maktura *et al.*, 2021; ). This strategy is still employed alone or in conjunction with other control agents to combat a variety of fruit fly species, despite the negative environmental effects of pesticide application. Dipterex, Malathion, Endrin, Dialordin, Diazinon, and Dimecron were used in Pakistan as a cover spray on mango orchards to prevent *B. zonata* growth.

Various pesticides target distinct fruit fly growth stages (Maula et al., 2022). While diazinon attacks emerging adults and popping larvae, malathion targets the adults. The lethal concentration of diazinon in males was 9.73 ppm while in females it was 14.12 ppm. According to (El-Gendy et al., 2021) diazinon was the most hazardous substance to B. zonata at 24 hours following treatment, followed by methoxy fenozide, and lufenuron, malathion. To suppress the population of fruit flies, diazinon has been widely applied as a soil drench. The practical implementation of chemicals has had many negative effects, including excessive insecticide residue in the environment. (Mahmood et al., 2016).

Li *et al.* (2021) evaluated the sub-lethal concentration of insecticides on the growth and development of *B. dorsalis*. Six insecticides were assessed against the progeny of *B. dorsalis* by using age stage, and two sex life tables. Insecticides negatively affect growth and development. The maximum reduction was observed in chlorpyrifos.

## **Conflicts of interest**

There is no conflict of interest related to this article.

#### Author's contribution

All authors have contributed equally

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### **References:**

- Abd-Elgawad, M.M. 2021. The mediterranean fruit fly (Diptera: Tephritidae), a key pest of citrus in Egypt. *J. Integr. Pest Manag.* 12: 28.
- Adly, D. 2016. Thermal Requirements of the Peach Fruit Fly, *Bactrocera zonata* (Saunders) (Diptera: Tephritidae), and its Exotic Parasitoid Species *Aganaspis daci* (Weld) (Hymenoptera: Eucoilidae). Egypt. Acad. *J. Biol. Sci. Entomol.*, 9: 89–96.
- Adnan, S.M., I. Farhana, J. Inskeep, P. Rempoulakis and P.W. Taylor. 2020. Dietary methoprene enhances sexual competitiveness of sterile male Queensland fruit flies in field cages. *J. Pest Sci.*, 93: 477–489.
- Afzal, M., and H. Javed. 2001. Evaluation of soaked wooden killer blocks for male annihilation (MA) on fruit fly *Bactrocera* Spp. (Diptera: Tephritidae). *Online J. Biol. Sci.*, 1: 577-579.
- Afzal, M., R. Mahmood and J. Stonehouse 2000. Soaked-wood killer blocks for mat fruit fly control in Pakistan. Proceedings of the Indian Ocean Commission, Regional Fruit Fly Symposium, Flic en Flac, Mauritius, 5th–9th June, 2000 Quatre Bornes: *Indian Ocean Commission*. p. 97100.
- Agarwal, M.L., K. Pramod and K. Vinod. 1999. Population suppression of *Bactrocera dorsalis* (Hendel) by *Bactrocera zonata* (Saunders)(Diptera: Tephritidae) in North Bihar. *Shashpa*, 6: 189–191.
- Ahmad, J.N., T. Sharif, S.J.N. Ahmad, S. Maqsood and F. Zafar. 2019. Molecular Identification and Characterization of Fruit Flies of Genus *Bactrocera* (Diptera: Tephritidae) in Pakistan. *Pak. J. Zool.*, 51.
- Ali, S.A.I., S.A. Mohamed, M.E. Mahmoud, S.A. Sabiel, S. Ali and A. Ali. 2014. Monitoring of Tephritidae of fruit trees and their level

- of infestation in South Kordofan State, Sudan. *Int. J. Agric. Innov. Res.*, 2: 2319–1473.
- Aluja, M. 2020. Future trends in fruit fly management. Fruit Fly Pests. CRC Press. pp.309–320.
- Amin, A.A. 2013. Efficiency of wooden blocks and cotton wicks as dispensers of methyl eugenol for attracting and kill the peach fruit fly males. *J. Plant Protect Pathol.* 4:245-252.
- Amin, A.A. 2017. Field and laboratory studies on infestation of immature mango fruits by the peach fruit fly, *Bactrocera zonata* (Saunders). *Egypt. J. Agric. Res.* 95:89–106
- Amin, M.R., R. Ara, S. Mukherjee, M.M.H. Bhuyain and M.N. Uddin. 2008. Economic evaluation of different management practices of cucurbit fruit fly (*Bactrocera cucurbitae*) in sweet gourd. *J. Sci. Technol.*, 6: 109–113.
- Anjum, S., M. Razaq, and M.S. Yazadni. 2000. Studies on seasonal activity and control of fruit flies (*Dacus* spp.) on mango (*Mangifera indica L.*) at Faisalabad, Pakistan. *Arab J. Plant Protect.*, 18: 121-123.
- Arif, M., M. Aasi, M. Farooq, H. Ali, S. Ul Islam, M. Asad, M. Shakeel and W. Zujian. 2017. Spatio-temporal distribution of the peach fruit fly, *Bactrocera zonata* (Diptera: Tephritidae) infesting citrus orchards at Sargodha, Pakistan. *Acta Entomol. Sin.*, 60: 1457–1466.
- Aryal, S., U.N. Nielsen, N.H. Sumaya, C. Wilson and M. Riegler. 2022. Virulence, penetration rate and reproductive potential of entomopathogenic nematodes from eastern Australia in Queensland fruit fly, Bactrocera tryoni. *Biol. Control*, 169: 104871.
- Bagheri, A., R. Kolyaee, M. Askari Seyahooei, S.S. Modaress Najafabadi and G. Faraji. 2017. Efficacy of methyl eugenol bait traps for controlling the mango fruit fly *Bactrocera zonata* (Diptera: Tephritidae). *J. Crop Prot.*, 6: 181–189.
- Baker, R., G. Gilioli, C. Behring, D. Candiani, A. Gogin, T. Kaluski, M. Kinkar, O. Mosbach-Schulz, F.M. Neri and S. Preti. 2019. *Bactrocera dorsalis* pest report to support ranking of EU candidate priority pests. *EN*-1641 44.

- Balagawi, S., S. Vijaysegaran, R.A. Drew and S. Raghu. 2005. Influence of fruit traits on oviposition preference and offspring performance of *Bactrocera tryoni* (Froggatt)(Diptera: Tephritidae) on three tomato (*Lycopersicon lycopersicum*) cultivars. *Aust. J. Entomol.* 44:97–103.
- Balmès, V. and R. Mouttet. 2017. Development and validation of a simplified morphological identification key for larvae of tephritid species most commonly intercepted at import in Europe. *EPPO Bull.* 47:91–99.
- Bayoumy, M. H., and M. M. El-Metwally. 2017. Daily flight activity rhythms of the peach and Mediterranean fruit flies using sexual and olfactory attractants. Acta Phytopathol. *Entomol. Hungarica*. 52:227-244.
- Ben-Yosef, M., E. Verykouki, Y. Altman, E. Nemni-Lavi, N.T. Papadopoulos and D. Nestel. 2021. Effects of Thermal Acclimation on the Tolerance of *Bactrocera zonata* (Diptera: Tephritidae) to Hydric Stress. *Front. Physiol.* 12:686424.
- Bertolini, E., C. Kistenpfennig, P. Menegazzi, A. Keller, M. Koukidou and C. Helfrich-Förster. 2018. The characterization of the circadian clock in the olive fly *Bactrocera oleae* (Diptera: Tephritidae) reveals a Drosophila-like organization. *Sci. Rep.* 8:1–12.
- Bestea, L., A. Réjaud, J.-C. Sandoz, J. Carcaud, M. Giurfa and M.G. de Brito Sanchez. 2021. Peripheral taste detection in honey bees: What do taste receptors respond to? *Eur. J. Neurosci.* 54:4417–4444.
- Binyameen, M., A. Hamid, I. Afzal, M. Sajjad, M. Azeem, S.M. Zaka, Z.M. Sarwar, S.A. Shad, T.C. Baker and F. Schlyter. 2021. Role of fruit volatiles of different guava varieties in attraction and oviposition behaviors of peach fruit fly, *Bactrocera zonata* Saunders. *Arthropod-Plant Interact.* 15:95–106.
- Boulahia-Kheder, S. 2021a. Review on major fruit flies (Diptera: Tephritidae) in North Africa: Bio-ecological traits and future trends. *Crop Prot.* 140:105416.
- Boulahia-Kheder, S. 2021b. Advancements in management of major fruit flies (Diptera: Tephritidae) in North Africa and future challenges: A review. *J. Appl. Entomol.* 145:939–957.

- Castro-Vargas, C., G. Pandey, H.L. Yeap, S.S. Prasad, M.J. Lacey, S.F. Lee, S.J. Park, P.W. Taylor and J.G. Oakeshott. 2023. Genetic variation for rectal gland volatiles among recently collected isofemale lines and a domesticated strain of Queensland fruit fly, *Bactrocera tryoni* (Diptera: Tephritidae). *Plos One* 18:e0285099.
- Charlery De La Masseliere, M., V. Ravigné, B. Facon, P. Lefeuvre, F. Massol, S. Quilici and P.-F. Duyck. 2017. Changes in phytophagous insect host ranges following the invasion of their community: Long-term data for fruit flies. *Ecol. Evol.* 7:5181–5190.
- Chiu, M.-C., Y.-H. Ni, K.-H. Lu and M.-H. Kuo. 2019. Effects of experimental warming on wild and transgenic oriental fruit fly, *Bactrocera dorsalis. J. Appl. Entomol.* 143:1022–1031.
- Choudhary, J.S., S.S. Mali, N. Naaz, D. Mukherjee, L. Moanaro, B. Das, A.K. Singh, M.S. Rao and B.P. Bhatt. 2020. Predicting the population growth potential of *Bactrocera zonata* (Saunders)(Diptera: Tephritidae) using temperature development growth models and their validation in fluctuating temperature condition. *Phytoparasitica* 48:1–13.
- Choudhary, J.S., S.S. Mali, N. Naaz, S. Malik, B. Das, A.K. Singh, M.S. Rao and B.P. Bhatt. 2021. Spatio and temporal variations in population abundance and distribution of peach fruit fly, *Bactrocera zonata* (Saunders) during future climate change scenarios based on temperature driven phenology model. Clim. *Risk Manag*. 32:100277.
- Courtney, S.P. and T.T. Kibota. 2017. Mother doesn't know best: selection of hosts by ovipositing insects. Insect-plant interactions. CRC press. pp.161–188.
- Darwish, A. A. E. 2014. Population density, diurnal activity and effect of trap height on capturing of peach fruit fly, *Bactrocera zonata* (saunders) and mediterranean fruit fly, *Ceratitis capitata* (wiedemann) in guava orchards at Nubaria region, *Egypt. J. Plant Protect. Pathol.* 5:89-98.
- de Oliveira, F.Q., J.B. Malaquias, W.R. de Souza Figueiredo, J. de Luna Batista and E.B. Beserra. 2012. Inhibition of fruit infestation by Mediterranean fruit fly using natural products. *Afr. J. Biotechnol*. 11:13922.

- de Souza, A.L.C., C.H.C. de Matos and R. Campos e Silva. 2022. Volatile Allelochemicals. Essential Oils: Applications and Trends in Food Science and Technology. Springer. pp.247–261.
- Deguine, J.-P., J.-N. Aubertot, R.J. Flor, F. Lescourret, K.A. Wyckhuys and A. Ratnadass. 2021. Integrated pest management: good intentions, hard realities. A review. *Agron. Sustain. Dev.* 41:38.
- Delrio, G. and A. Cocco. 2012. peach fruit fly, Bactrocera zonata: a major threat for Mediterranean fruit crops? Acta Hortic.
- Dias, N.P., M.J. Zotti, P. Montoya, I.R. Carvalho and D.E. Nava. 2018. Fruit fly management research: A systematic review of monitoring and control tactics in the world. *Crop Prot.* 112:187–200.
- Draz, K.A. 2016. Population activity of peach fruit fly *Bactrocera zonata* (Saunders)(Diptera: Tephiritidae) at fruits orchards in Kafer El-Shikh Governorate, Egypt. *Arthropods* 5:28.
- Drew, R.A.I. 1987. Behavioural strategies of fruit flies of the genus *Dacus* (Diptera: Tephritidae) significant in mating and hostplant relationships. *Bull. Entomol. Res.* 77:73–81.
- Drummond, F., E. Ballman and J. Collins. 2019.

  Population Dynamics of Spotted Wing
  Drosophila (*Drosophila suzukii*(Matsumura) in Maine Wild Blueberry
  (*Vaccinium angustifolium* Aiton). *Insects*10:205.
- Duyck, P.-F., H. Jourdan and C. Mille. 2022. Sequential invasions by fruit flies (Diptera: Tephritidae) in Pacific and Indian Ocean islands: A systematic review. *Ecol. Evol.* 12:8880.
- El-Gendy, I.R., M.I. El-Banobi and J.A. Villanueva-Jimenez. 2021. Bio-pesticides alternative diazinon to control peach fruit fly, *Bactrocera zonata* (Saunders)(Diptera: Tephritidae). *Egypt. J. Biol. Pest Control* 31:1–8.
- Elnagar, S., M. El-Sheikh, A. Hashem, and Y. Afia, 2010. Recent invasion by *Bactrocera zonata* (Saunders) as a new pest competing with *Ceratitis capitata* (Wiedemann) in attacking fruits in Egypt. *Asp. Appl. Biol.* 104: 97-102.

- Gadenne, C., R.B. Barrozo and S. Anton. 2016.

  Plasticity in Insect Olfaction: To Smell or
  Not to Smell? *Annu. Rev. Entomol.*61:317–333.
- Ghanim, N. M. 2009. Studies on the peach fruit fly, Bactrocera zonata (Saunders) (Tephritidae, Diptera). Unpublished Ph. D. Thesis, Fac. Agric., Mansoura Univ.
- Ghanim, N. M. 2013. Influence of methyl eugenol diluted with paraffin oil on male annihilation technique of peach fruit fly, *Bactrocera zonata* (Saunders) (Diptera: Tephritidae). *Entomol. Ornithol. Herpetol.* 2: 1-6.
- Grechi, I., A. L. Preterre, A. Caillat, F. Chiroleu, and A. Ratnadass. 2021. Linking mango infestation by fruit flies to fruit maturity and fly pressure: A prerequisite to improve fruit fly damage management via harvest timing optimization. *Crop Protection.* 146:105-663.
- Gupta, D., and R. Bhatia. 2000. Population fluctuations of fruit flies, *Bactrocera* spp. in sub mountainous mango and guava orchards. *J. Appl. Horti.* 2:47-49.
- Hasnain, M., S. Saeed, U. Naeem-Ullah, S. Ullah, and S. Zaka, 2023. Synergist Response of the Peach fruit fly, *Bactrocera zonata* (Sunders) to some ammonium based proteinaceous food bait attractants.
- He, Y., Y. Xu and X. Chen. 2023. Biology, Ecology and Management of Tephritid Fruit Flies in China: A Review. Insects 14:196.
- Hee, A.K.-W. and K.-H. Tan. 2006. Transport of methyl eugenol-derived sex pheromonal components in the male fruit fly, *Bactrocera dorsalis*. Comp. *Biochem. Physiol. Part C Toxicol. Pharmacol.* 143:422–428.
- Hosseini, S.A., S.H. Goldansaz, S.B. Menken, M. van Wijk, P. Roessingh and A.T. Groot. 2017. Field attraction of carob moth to host plants and conspecific females. *J. Econ. Entomol.* 110:2076–2083.
- Inskeep, J.R. 2019. The behaviour of the Queensland fruit fly, *Bactrocera tryoni* (Diptera: Tephritidae), and its implications for improving integrated pest management using the sterile insect technique. PhD Thesis, Macquarie University.
- Jaworski, T. and J. Hilszczański. 2013. The effect of temperature and humidity changes on insects development their impact on forest

- ecosystems in the context of expected climate change.
- Jin, S., X. Zhou, F. Gu, G. Zhong, and X. Yi, 2017.

  Olfactory plasticity: variation in the expression of chemosensory receptors in *Bactrocera dorsalis* in different physiological states. Frontiers *Physiol*. 8:672.
- Joachim-Bravo, I.S., O.A. Fernandes, S.A. Bortoli and F.S. Zucoloto. 2001. Oviposition behavior of *Ceratitis capitata* Wiedemann (Diptera: Tephritidae): association between oviposition preference and larval performance in individual females. *Neotrop. Entomol.* 30:559–564.
- Kapoor, V. 1986. Ml agarwal. Fruit Flies Econ. Importance 84 9647:51.
- Kausar, A., F. Ullah, F. Jahan, K. Khan, S. Wahid, G. Tanzila and N.H. Khan. 2022. Bionomics of *Bactrocera* fruit flies (Diptera: Tephritidae) in Khyber Pakhtunkhwa, Pakistan; exploring performance of various trap types and their characteristics. *Fla. Entomol.*, 105:231–242.
- Kay, B.J. 2023. The complexity of competition between three native Australian fruit fly species, *Bactrocera tryoni*, *B. neohumeralis* and *B. jarvisi* in a changing environment. PhD Thesis, Queensland University of Technology.
- Khan, I., A.S. Khan, I. Rajwana, A. Khan, M. Abubakkar and S. Raza. 2016. Premium Quality Mango Genotypes for Extended Harvest Season. *HortScience*, 51:1609–1612.
- Khan, R.A. and M. Naveed. 2017. Evaluation of comparative toxicity of different insecticides against fruit fly, *Bactrocera zonata* Saunders (Diptera: Tephritidae). *Pak. J. Zool.*, 49.
- Khosravi, M., N. Sahebzadeh, R. Kolyaie, and A. Mokhtari. 2018. Field evaluation of controlling methods of mango fruit flies *Bactrocera zonata* (Diptera: Tephritidae) in the southern part of Iran. *Trakia J. Sci.*, 1: 62-69.
- Koswanudin, D., A. Basukriadi, I.M. Samudra and R. Ubaidillah. 2018. Host preference fruit flies *Bactrocera carambolae* (Drew & Hancock) and *Bactrocera dorsalis* (Drew and Hancock) (Diptera: Tephritidae). *J. Entomol. Indones*. 15:40–40.

- Kumar, L., J.S. Choudhary and B. Kumar. 2021.

  Host plant-mediated effects of elevated CO2 and temperature on growth and developmental parameters of *Zygogramma bicolorata* (Coleoptera: Chrysomelidae).

  Bull. Entomol. Res. 111:111–119.
- Levi-Zada, A., A. Levy, P. Rempoulakis, D. Fefer, S. Steiner, Y. Gazit, D. Nestel, B. Yuval and J.A. Byers. 2020. Diel rhythm of volatile emissions of males and females of the peach fruit fly *Bactrocera zonata*. J. *Insect Physiol*. 120:103970.
- Liendo, M., A. Parreño, A. Pietrek, J. Bouvet, F. Milla, T. Vera, J. Cladera and D. Segura. 2020. Infestation of fruit by conspecific and heterospecific females deters oviposition in two Tephritidae fruit fly species. *J. Appl. Entomol.*, 144.
- Lunau, K. 2014. Visual ecology of flies with particular reference to colour vision and colour preferences. *J. Comp. Physiol. A Neuroethol. Sens. Neural. Behav. Physiol.*, 200: 497–512.
- Lux, S.A., K. Michalska, A. Wnuk, D. Paliczuk, H. Vogt, T. Hernik and M. Studnicki. 2017. Female age as a factor determining the patterns of tree canopy utilisation by the European cherry fruit fly, *Rhagoletis cerasi. Int. J. Pest Manag.*, 63:371–381.
- Mahmood, I., S.R. Imadi, K. Shazadi, A. Gul and K.R. Hakeem. 2016. Effects of pesticides on environment. Plant Soil Microbes. *Implic. Crop Sci.*, 1: 253–269.
- Mahmoud, M.E.E., S.A. Mohamed, F.M. Khamis, M.A.I. Basher and S. Ekesi. 2019. The role of *Tetrastichus giffardianus* Silvestri (Eulophidae: Hymenoptera) in natural control of *Bactrocera zonata* (Saunders) (Tephritidae: Diptera) and its temporal abundance in Sudan. EPPO Bull. 49:359–363.
- Mandanayake, M.A., S. Shohaimi, N.I. Ghani and A.K. Hee. 2023. Establishment of nonmethyl eugenol-responding lines from feral Oriental fruit fly, *Bactrocera dorsalis* (Hendel) (Diptera Tephritidae). Phytoparasitica 1–12.
- Manoukis, N.C., E.B. Jang and R.V. Dowell. 2017. Survivorship of male and female *Bactrocera dorsalis* in the field and the effect of male annihilation technique. *Entomol. Exp. Appl.* 162:243–250.
- Manoukis, N.C., R.I. Vargas, L. Carvalho, T. Fezza, S. Wilson, T. Collier and T.E. Shelly. 2019.

- A field test on the effectiveness of male annihilation technique against *Bactrocera dorsalis* (Diptera: Tephritidae) at varying application densities. *PLOS ONE* 14:0213337.
- Manrakhan, A. 2016. Detection and monitoring of fruit flies in Africa. Fruit Fly Res. Dev. Afr.- Sustain. Manag. *Strategy Improve Hortic.* 253–273.
- Manrakhan, A., J.-H. Daneel, R. Beck, M. Virgilio, K. Meganck and M. De Meyer. 2017. Efficacy of trapping systems for monitoring of Afrotropical fruit flies. *J. Appl. Entomol.* 141:825–840.
- Mariano, D.O.C., Ú.C. de Oliveira, A.J. Zaharenko, D.C. Pimenta, G. Rádis-Baptista and Á.R. de Brandão Prieto-da. 2019. Bottom-up proteomic analysis of polypeptide venom components of the giant ant Dinoponera quadriceps. *Toxins* 11.
- Marwat, S.K., K. Usman, N. Khan, M.U. Khan, E.A. Khan, M.A. Khan and A. ur Rehman. 2013. Weeds of wheat crop and their control strategies in Dera Ismail Khan district, Khyber Pakhtun Khwa, Pakistan.
- Maula, F., A.A. Khan, A. Ali, I., M. Younus, M. Israr, M.A. Rauf and I.A. Khan. 2022. Evaluation of different traps and lures combinations for monitoring and ecofriendly management of fruit fly (*Bactrocera spp*) in peach orchards. J. *Entomol. Zool. Stud.*, 10:105–110.
- Momen, F.M., A.-E.K. Nasr, A.-E.M. Metwally, Y.A. Mahmoud and K.M. Saleh. 2016. Performance of five species of phytoseiid mites (Acari: Phytoseiidae) on *Bactrocera zonata* eggs (Diptera: Tephritidae) as a factitious food. *Acta Phytopathol. Entomol. Hung.* 51:123–132.
- Murtaza, G., F. Ullah, T. Ahmad, M. Ramzan, M. Daud, S. Iqbal and A. Ali. 2021. Biology and morphology of *Bactrocera dorsalis* and *Bactrocera zonata* on guava under laboratory conditions. 180–185.
- Muthuthantri, S. and A.R. Clarke. 2012. Five commercial citrus rate poorly as hosts of the polyphagous fruit fly *Bactrocera tryoni* (Froggatt) (Diptera: T ephritidae) in laboratory studies. *Aust. J. Entomol.*, 51:289–298.
- Myers, S.W., E. Cancio-Martinez, G.J. Hallman, E.A. Fontenot and M.J. Vreysen. 2016. Relative tolerance of six *Bactrocera* (Diptera: Tephritidae) species to

- phytosanitary cold treatment. J. Econ. Entomol. 109:2341–2347.
- Navarro-Llopis, V. and S. Vacas. 2014. Mass trapping for fruit fly control. Trapp. Detect. Control Regul. Tephritid Fruit Flies Lures Area-Wide Programs *Trade Implic.* 513–555.
- Ndlela, S., A.G. Azrag and S.A. Mohamed. 2021. Determination of temperature thresholds for the parasitoid *Diachasmimorpha longicaudata* (Hymenoptera: Braconidae), using life cycle simulation modeling: Implications for effective field releases in classical biological control of fruit flies. *Plos One* 16:0255582.
- Ndzana Abanda, R.F.X., T. Turlings, N. Woin and S. Quilici. 2016. Factors influencing the mating success of *Bactrocera zonata* (Saunders) (Diptera: Tephritidae) males in an sit programme. Tephritid Workers of Europe, Africa and the Middle East: 3rd International Symposium 11-14 April 2016 Stellenbosch, South Africa. Available at https://agritrop.cirad.fr/583108/.
- Noman, Q.M., F.M. Shah, K. Mahmood and M. Razaq. 2021. Population Dynamics of Tephritid Fruit Flies in Citrus and Mango Orchards of Multan, Southern Punjab, Pakistan. *Pak. J. Zool.*, 54.
- Paranhos, B.A., S. Poncio, R. Morelli, D.E. Nava, L.A. de Sá and N.C. Manoukis. 2021. Nontarget effects of the exotic generalist parasitoid wasp *Fopius arisanus* (Sonan) estimated via competition assays against *Doryctobracon areolatus* (Szepligeti) on both native and exotic fruit fly hosts. *BioControl.*, 66: 83–96.
- Paranhos, B.J., D.E. Nava and A. Malavasi. 2019. Biological control of fruit flies in Brazil. *Pesqui. Agropecuária Bras.* 54:26037.
- Perry, R. N., R. U. Ehlers, and I. Glazer. 2012. A realistic appraisal of methods to enhance desiccation tolerance of entomopathogenic nematodes. *J. nematol.* 44:185.
- Petitinga, C.S.C.D., A.K.P. Roriz and I.S. Joachim-Bravo. 2021. Competitive strategies between *Anastrepha fraterculus* and *Anastrepha obliqua* (Diptera: Tephritidae) regulating the use of host fruits. *J. Appl. Entomol.*, 145:612–620.
- Prakash, S. and Y. Kaur. 2022. Sustainable Development Goals and 5P'S. *Environ. Sustain.* 4:116.

- Quilici, S., T. Atiama-Nurbel and T. Brévault. 2014. Plant odors as fruit fly attractants. *Trapp. Detect. Control Regul. Tephritid Fruit Flies Lures Area-Wide Programs Trade Implic.* 119–144.
- Ragab, S. K., and N. M. Youssef. 2021. Effect of blending ammonium acetate and diammonium phosphate solutions on their attractance to Mediterranean fruit fly, *Ceratitis capitata* in mandarin orchids under field conditions. *world* (Liquido *et al.*, 1991; White and Elson-Harris, 1992; *Papadopoulos*, 20:25.
- Rauf, I., A. Nazir, M. S, I. Muhammad and M. Khan. 2013. Laboratory studies on ovipositional preference of the peach fruit fly *Bactrocera zonata* (Saunders) (Diptera: Tephiritidae) for different host fruits. *Afr. J. Agric. Res.*, 8:1300–1303.
- Saafan, M.H. 2005. Field evaluation of some attractants for attracting the adults of mediterranean fruit fly, *Ceratitis capitata* (Wiedemann) and peach fruit fly, *Bactrocera zonata* (saunders) in citrus orchards. *Egypt. J. Agric. Res.*, 83: 1141–1156.
- Saeed, M., T. Ahmad, M. Alam, L.A. Al-Shuraym, N. Ahmed, M. Ali Alshehri, H. Ullah and S.M. Sayed. 2022. Preference and performance of peach fruit fly (*Bactrocera zonata*) and Melon fruit fly (*Bactrocera cucurbitae*) under laboratory conditions. *Saudi J. Biol. Sci.*, 29:2402–2408
- Saleh, M., N. El-Wakeil, H. Elbehery, N. Gaafar, and S. Fahim. 2019. Biological pest control for sustainable agriculture in Egypt. Sustain Agri Environ Egypt: Part II: Soil-Water-Plant Nexus, 145-188.
- Salgado, A.L. and M. Saastamoinen. 2019. Developmental stage-dependent response and preference for host plant quality in an insect herbivore. *Anim. Behav.*, 150:27–38.
- Salman, M., M.H. Khan, M. Zahid, G.Z. Khan, F. Rahim and U. Khalique. 2022. Comparative Ovipositional Preference of the Peach Fruit Fly, *Bactrocera zonata* (Saunders)(Diptera: Tephritidae) for Selected Fruits under Free-Choice Laboratory Setting. *Sarhad J. Agric.*, 38:572–577.
- Sarwar, M., M. Hamed, B. Rasool, M. Yousaf and M. Hussain. 2013. Host preference and performance of fruit flies *Bactrocera zonata* (Saunders) and *Bactrocera*

- cucurbitae (Coquillett)(Diptera: Tephritidae) for various fruits and vegetables. *Int. J. Sci. Res. Environ. Sci.*, 1:188–194.
- Sharma, R. and V.R.R. Sanikommu. 2017.

  Preharvest fruit bagging for better protection and postharvest quality of horticultural produce. *Preharvest Modulation of Postharvest Fruit and Vegetable Quality*. 455–489.
- Shaurub, E.-S.H. 2023. Review of entomopathogenic fungi and nematodes as biological control agents of tephritid fruit flies: current status and a future vision. *Entomol. Exp. Appl.*, 171:17–34.
- Sidahmed, O. A., A. K. Taha, G. A. Hassan, and I. F. Abdalla. 2014. Evaluation of pheromone dispenser units in methyl eugenol trap against *Bactrocera invadens* Drew, Tsuruta and White (Diptera: Tephritidae) in Sudan. *Sky J. Agri. Res.*, 3:148-151
- Stanley, J., J. P. Gupta, and D. Rai. 2015. Population dynamics of fruit flies, *Bactrocera* spp. in northwestern Himalaya. *Ind. J. Entomol.*, 77:214-220.
- Stejskal, V., T. Vendl, R. Aulicky, and C. Athanassiou. 2021. Synthetic and natural insecticides: Gas, liquid, gel and solid formulations for stored-product and foodindustry pest control. *Insects.* 12: 590.
- Tangpao, T., N. Phuangsaujai, S. Kittiwachana, D.
  R. George, P. Krutmuang, B. Chuttong, and S. R. Sommano. 2022. Evaluation of Markers Associated with Physiological and Biochemical Traits during Storage of 'Nam Dok Mai Si Thong' Mango Fruits. Agri. 12:1407.
- Ugwu, J.A., O.Y. Alabi and O.J. Aluko. 2021. Insecticidal activity of crude extracts of three spices and commercial botanical pesticide on oriental fruit fly under laboratory conditions. *J. Basic Appl. Zool.*, 82:26.
- Ullah, F., H. Gul, A. Güncan, M. Hafeez, K. Tariq, N. Desneux and Z. Li. 2022. Short-Term Temperature Stress Modulates Fitness Traits in *Bactrocera zonata*, through Negative Impact on Larval Stage. *Agronomy* 12:2903.
- Valero-Jiménez, C.A., H. Wiegers, B.J. Zwaan, C.J.M. Koenraadt and J.A.L. van Kan. 2016. Genes involved in virulence of the entomopathogenic fungus *Beauveria bassiana*. *J. Invertebr. Pathol.*, 133: 41–49.

- Vargas, P. Jc and L. L. 2015. An overview of pest species of *Bactrocera* fruit flies (Diptera: Tephritidae) and the integration of biopesticides with other biological approaches for their management with a focus on the pacific region. *Insects* 6.
- Wei, Y., L. Li, S. Pan, Z. Liu, J. Fan, and M. Tang, 2023. Adaptive reproductive strategies of an *Ectoparasitoid Sclerodermus* guani under the stress of its entomopathogenic fungus *Beauveria bassiana*. *Insects.* 14: 320.
- Yesmin, M. 2019. Bio-ecology host preference and management of cucurbit fruit FLY. PhD Thesis, Department of Entomology.
- Zain-Ul-Aabdin Abro, N.B., R.M. Memon, N.H. Khuhro and Q.A. Soomro. 2020. 89. Population variations of fruit flies, *Bactrocera* spp. in mango orchards of Hyderabad and Larkana Sindh. *Pure Appl. Biol. PAB* 9:949–955.